# OPPICEC

# IUCLID

## **Data Set**

**Existing Chemical** 

CAS No.

EINECS Name

EC No.

TSCA Name Molecular Formula : ID: 107-12-0

: 107-12-0

: propiononitrile : 203-464-4

: Propanenitrile

: C3H5N

Producer related part

Company Creation date : Solutia Inc. : 06.06.2003

Substance related part

Company

Creation date

: Solutia Inc.

: 06.06.2003

Status Memo

•

Printing date Revision date : 06.10.2003 : 07.06.2004

Date of last update

: 30.10.2003

Number of pages

: 58

Chapter (profile) Reliability (profile) : Chapter: 1, 2, 3, 4, 5, 6, 7, 8, 10 : Reliability: without reliability, 1, 2, 3, 4

Flags (profile)

: Flags: without flag, confidential, non confidential, WGK (DE), TA-Luft (DE),

Material Safety Dataset, Risk Assessment, Directive 67/548/EEC, SIDS

#### 1. General Information

**Id** 107-12-0 Date 02.10.2003

#### 1.0.1 APPLICANT AND COMPANY INFORMATION

#### 1.0.2 LOCATION OF PRODUCTION SITE, IMPORTER OR FORMULATOR

#### 1.0.3 IDENTITY OF RECIPIENTS

#### 1.0.4 DETAILS ON CATEGORY/TEMPLATE

#### 1.1.0 SUBSTANCE IDENTIFICATION

**IUPAC Name** 

Smiles Code : C(#N)CC
Molecular formula : C3H5N
Molecular weight : 55.079
Petrol class

Petrol class

11.06.2003

#### 1.1.1 GENERAL SUBSTANCE INFORMATION

Purity type : typical for marketed substance

Substance type : organic
Physical status : liquid
Purity : = 99.6 : = 99.6 % v/v Colour : clear Odour : pungent

11.06.2003 (31)

#### 1.1.2 SPECTRA

#### 1.2 **SYNONYMS AND TRADENAMES**

#### cyanoethane

10.07.2003

ethyl cyanide

10.07.2003

propanenitrile

10.07.2003

#### propionic nitrile

10.07.2003

## **Id** 107-12-0 1. General Information **Date** 02.10.2003 propionitrile 10.07.2003 propylnitrile 10.07.2003 1.3 IMPURITIES 1.4 ADDITIVES 1.5 TOTAL QUANTITY 1.6.1 LABELLING 1.6.2 CLASSIFICATION 1.6.3 PACKAGING 1.7 USE PATTERN : industrial Type of use Category Reliability : (2) valid with restrictions 11.06.2003 1.7.1 DETAILED USE PATTERN 1.7.2 METHODS OF MANUFACTURE **REGULATORY MEASURES** 1.8

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1.8.1 OCCUPATIONAL EXPOSURE LIMIT VALUES

1.8.2 ACCEPTABLE RESIDUES LEVELS

#### 1. General Information

ld 107-12-0 **Date** 02.10.2003

1.8.3 WATER POLLUTION

1.8.4 MAJOR ACCIDENT HAZARDS

1.8.5 AIR POLLUTION

#### 1.8.6 LISTINGS E.G. CHEMICAL INVENTORIES

Type : EINECS

Additional information :

11.06.2003

Type : TSCA

Additional information :

11.06.2003

#### 1.9.1 DEGRADATION/TRANSFORMATION PRODUCTS

1.9.2 COMPONENTS

1.10 SOURCE OF EXPOSURE

1.11 ADDITIONAL REMARKS

1.12 LAST LITERATURE SEARCH

1.13 REVIEWS

ld 107-12-0 **Date** 02.10.2003

#### 2.1 MELTING POINT

Value : = -92.8 °C

Sublimation

Method : other

Year

GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

Remark : Purity of material is unknown.
Reliability : (2) valid with restrictions

Source is peer-reviewed, published data.

Flag : Critical study for SIDS endpoint

11.06.2003 (28)

#### 2.2 BOILING POINT

**Value** : = 97 °C at 1013 hPa

Decomposition

Method : other: not specified

Year

GLP : no data

**Test substance** : as prescribed by 1.1 - 1.4

Remark : Purity of material is unknown.
Reliability : (2) valid with restrictions

Source is peer-reviewed, published data.

Flag : Critical study for SIDS endpoint

10.07.2003 (28)

#### 2.3 DENSITY

Type : relative density

Value : = .7818 g/cm³ at 20 °C

Method : other: not specified

Year

GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

Remark : Purity of material is unknown.
Reliability : (2) valid with restrictions

Source is peer-reviewed, published data.

10.07.2003 (36)

#### 2.3.1 GRANULOMETRY

#### 2.4 VAPOUR PRESSURE

**Value** : = 52 hPa at 20 °C

Decomposition

Method : other (measured)

Year :

GLP : no

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ld 107-12-0 **Date** 02.10.2003

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: Purity of the test material is 99.6%.

**Reliability** : (2) valid with restrictions

Source of data is a MSDS.

Flag : Critical study for SIDS endpoint

29.07.2003 (31)

**Value** : = 53.3 at 22 °C

Decomposition

Method : other (measured): not specified

Year :

GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: Purity of the material is unknown.

**Result**: Value is 40 mm Hg, which converts to 53.3 hPa.

**Source**: Hazardous Substances Data Bank for Priopionitrile, dated 5/13/99.

**Reliability** : (2) valid with restrictions

Primary source is peer reviewed and published.

10.07.2003 (10)

#### 2.5 PARTITION COEFFICIENT

Partition coefficient :

**Log pow** : = .16 at °C

pH value

**Method** : other (measured)

Year

GLP : no data

**Test substance** : as prescribed by 1.1 - 1.4

**Test condition**: The value was obtained using the Shake-Flask method. The agueous

phase was octanol-saturated water. The concentration of material in the

aqueous phase was measured using gas-liquid chromatography.

**Test substance**: Purity of the test material was not mentioned.

**Reliability** : (2) valid with restrictions

Data were from a peer reviewed, published source.

Flag : Critical study for SIDS endpoint

13.08.2003 (30)

Partition coefficient : octanol-water Log pow : = .35 at 20 °C

pH value

Method : other (calculated)

Year : 2003 GLP : no

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: Measured inputs to the program are CAS No., melting point (-92.8 degrees

C), boiling point (97 degrees C), vapor pressure (39 mm Hg), and water

solubility (93380 mg/l).

**Reliability** : (2) valid with restrictions

Data were obtained by modeling.

13.08.2003 (17)

#### 2.6.1 SOLUBILITY IN DIFFERENT MEDIA

**Id** 107-12-0 **Date** 02.10.2003

Solubility in water

Value = 93380 mg/l at 25 °C

pH value

at °C concentration

Temperature effects

Examine different pol.

pKa at 25 °C

Description

Stable

Deg. product

Method other: not specified

Year

**GLP** no data

as prescribed by 1.1 - 1.4 **Test substance** 

Test substance Purity of the test material was not mentioned.

Reliability (2) valid with restrictions

Data were from a peer reviewed, published source.

: Critical study for SIDS endpoint Flag

13.08.2003 (37)

Solubility in water

Value = 119 g/l at 40 °C

pH value

concentration at °C

Temperature effects

Examine different pol.

pKa at 25 °C

Description

Stable

Deg. product

Method other: not specified

Year

**GLP** no data

as prescribed by 1.1 - 1.4 Test substance

Remark Purity of material is unknown.

Reliability (2) valid with restrictions Source is peer-reviewed, published data.

15.08.2003 (36)

Solubility in water

Value = 55650 mg/l at °C

pH value

concentration at °C

Temperature effects

Examine different pol.

pKa at 25 °C

Description Stable

Deg. product

Method other: calculated

Year 2003 **GLP** no

**Test substance** as prescribed by 1.1 - 1.4

Remark Measured inputs to the program are CAS No., melting point (-92.8 degrees

C), boiling point (97 degrees C), vapor pressure (39 mm Hg), and water

solubility (93380 mg/l).

Reliability (2) valid with restrictions

Data were obtained by modeling.

15.08.2003 (18)

ld 107-12-0 **Date** 02.10.2003

#### 2.6.2 SURFACE TENSION

#### 2.7 FLASH POINT

Value :  $= 16 \,^{\circ}\text{C}$ Type : open cup

Method : other: not specified

Year

GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: Purity of the test material is unknown.

**Source**: Hazardous Substances Data Bank for propionitrile, dated 5/13/99.

**Reliability** : (2) valid with restrictions

Primary reference is peer-reviewed and published.

10.07.2003 (23)

#### 2.8 AUTO FLAMMABILITY

#### 2.9 FLAMMABILITY

#### 2.10 EXPLOSIVE PROPERTIES

#### 2.11 OXIDIZING PROPERTIES

#### 2.12 DISSOCIATION CONSTANT

#### 2.13 VISCOSITY

#### 2.14 ADDITIONAL REMARKS

ld 107-12-0 **Date** 02.10.2003

#### 3.1.1 PHOTODEGRADATION

Type : air
Light source : Sun light
Light spectrum : nm

Relative intensity : based on intensity of sunlight

INDIRECT PHOTOLYSIS

Sensitizer : OH

Conc. of sensitizer

**Rate constant** : = .00000000001938 cm³/(molecule\*sec)

**Degradation** : = 50 % after 55.2 day(s)

Deg. product

Method : other (calculated)

Year : 2003 GLP : no

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: Measured inputs to the program are CAS No., melting point (-92.8 degrees

C), boiling point (97 degrees C), vapor pressure (39 mm Hg), and water

solubility (93,380 mg/l).

Reliability : (2) valid with restrictions

Data were obtained by modeling.

Flag : Critical study for SIDS endpoint

15.08.2003 (13)

#### 3.1.2 STABILITY IN WATER

Type : abiotic
Method : other
Year : 2003
GLP : no

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: EPIWIN Hydrowin cannot calculate hydrolysis rate constants for nitriles.

The theoretical hydrolysis of propionitrile and several other chemicals has been examined by Dr. Lee Wolfe at the USEPA Environmental Research Laboratory in Athens, Georgia. The results of these analyses were published in a report by Dr. Wolfe that could not be located. In a personal communication, Dr. Wolfe stated that propionitrile can hydrolyze (albeit slowly). According to a study cited in the Hazardous Substances Data Bank, the chemical hydrolysis of the related material acetonitrile in water is base-catalyzed (the rate constant for base catalyzed hydrolysis is 5.8X10-3/M-hr), but the half-life at pH 7 is more than 150,000 yrs (Ellington et al., 1988). Acetonitrile (CH₃C≡N, CAS No. 75-05-8) is the 2-carbon analog of the category members, possessing the same functionality, but having one less carbon than propionitrile. Taken together, these data suggest that hydrolysis of propionitrile at environmentally relevant pHs will occur too slowly to be a significant means of degradation.

Reliability : (2) valid with restrictions

Experimental results for the test material could not be located. Results are

for a related material.

07.08.2003 (16)

#### 3.1.3 STABILITY IN SOIL

ld 107-12-0 **Date** 02.10.2003

#### 3.2.1 MONITORING DATA

#### 3.2.2 FIELD STUDIES

#### 3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

Type : fugacity model level III

Media: other: air, water, soil and sedimentAir: 14.4 % (Fugacity Model Level I)Water: 48.7 % (Fugacity Model Level I)Biota: .0821 % (Fugacity Model Level II/III)Soil: 36.9 % (Fugacity Model Level II/III)

Method : other Year : 2003

**Remark**: Measured inputs to the program are CAS No., melting point (-92.8 degrees

C), boiling point (97 degrees C), vapor pressure (39 mm Hg), and water

solubility (93,380 mg/l).

Result : Henry's Law Constant (Bond Est) is estimated to be 4.06E-005 atm-

m3/mole. The soil/sediment constant Koc is 8.3 as estimated by the

EPIWIN PCKOC Program (v1.66).

Reliability : (2) valid with restrictions

Data were obtained by modeling.

Flag : Critical study for SIDS endpoint

15.08.2003 (15)

#### 3.3.2 DISTRIBUTION

#### 3.4 MODE OF DEGRADATION IN ACTUAL USE

#### 3.5 BIODEGRADATION

Type : aerobic

inoculum : other: activated sludge

Concentration : 1000 mg/l related to COD (Chemical Oxygen Demand)

Contact time : 6 hour(s)

**Result** : other: biodegradable

Deg. product

Method : other Year : 1960 GLP : no

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: The test conditions state that the test was performed for 6 hours, but the

results for 10 hours are listed.

The study results show that biodegradation occurred, but the results are not related to TOD. Based on the TOD of the previous study (1670 mg/l), the percentage of material that biodegraded in 10 hours was approximately

24%.

**Result** : The oxygen uptake was as follows:

Time (hrs) Approximate O2 uptake (mg/l)

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2	100
4	210
6	300
8	350
10	400

The study results show that 400 mg/l O2 out of a possible 1000 mg/l (COD) was utilized (40%).

The kinetics of the metabolism of propionamide and propionic acid suggested that propionitrile was sequentially metabolized to these materials.

#### **Test condition**

Bacteria: Activated sludges were grown in 1.5 liter pilot plants under 23 hour aeration. The material was allowed to settle for 1 hour. After settling, 1.0 liter of supernatant liquor was withdrawn, 1.0 liter of tap water, organics and buffer were added, and aeration was resumed. This cycle was repeated 7 days a week for the duration of the study.

Test material: The quantity of feed was 1000 mg/l (based on COD). Occasionally, at the end of the aeration period, 25 ml of mixed liquor was passed through a membrane filter, which was then dried at 103 degrees C and reweighed to determine the amount of suspended solids. The filter was then analyzed for COD and all forms of inorganic nitrogen.

Washed sludge, grown on propionitrile, was placed in a Warburg Respirometer with low concentrations of several possible breakdown products. Oxygen uptake was then measured for 6 hours. After this time. the contents of each flask were passed through a membrane filter so that the filtrate could be analyzed for COD and inorganic nitrogen forms.

Reliability

(2) valid with restrictions

Purity of the test material was not given.

Flag

: Critical study for SIDS endpoint

10.08.2003 (33)

Type : aerobic

Inoculum other: activated sludge

: 500 mg/l Concentration : 72 hour(s) Contact time

 $= 0 (\pm) \%$  after 72 hour(s) Degradation other: material was toxic Result

Deg. product

: other Method 1970 Year **GLP** : no

**Test substance** : as prescribed by 1.1 - 1.4

Result

: TOD for propionitrile was 1670 mg/l. Test material was toxic to all three

sludges. No biodegradation occurred.

**Test condition** 

: Bacteria: The activated sludges were obtained from the municipal plant at Franklin, TN, the municipal plant at Nashville TN and the plant at Bordeaux, a suburb of Nashville. Mixed liquor from the aeration tanks was collected in the morning, the day of the Warburg run. Each sample was packed in ice and transported to the laboratory within 1 hour of collection. Before the run began, the sludge sample was blended for 10 sec and the homogenous blend was analyzed for concentration of SS (not defined, but assumed suspended solids), using a membrane-filter technique. The original sample was adjusted to a SS concentration of 2,500 mg/l.

Test conduct: Test material was added to a Warburg flask (125 ml) in order to obtain a final concentration of 500 mg/l in the reaction compartment (final volume of 20 ml). KOH (1.0 ml, 20%) was added to the

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center well. A 20 ml volume of activated sludge was then introduced. The test was performed in duplicate. Flasks were incubated for 72 hours (constant motion) at 20 degrees C. All three sludges were tested. Oxygen uptake curves were plotted. Respiration of the sludge alone was plotted as the control curve.

Theoretical O2 demand (the mg/l O2 required to completely oxidize the test material) was calculated on the basis of the test material to CO2 and water, plus nitrate according to the following equation: [TOD = moles of O2 required to balance the equation x molecular weight of O2 x concentration of test material/ (moles of test material required to balance the oxidation equation x molecular weight of the test material)]. The percentage of TOD was to be calculated as follows: % TOD =  $100 \times D$  (the difference in mg/l of O2 uptake between substrate and control)/ TOD. The material was considered toxic if D was less than 0.

**Reliability** : (2) valid with restrictions

Purity of the test material was not given.

08.08.2003 (26)

Type : aerobic

**Inoculum** : other: mixed microbial culture

Concentration : 1000 mg/l Contact time : 48 hour(s)

Result : other: biodegradable

Deg. product

Method : other Year : 1992 GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

**Result**: The final protein and ammonia concentrations and pH were 10.20 mg/l,

76.9 micromoles/ml and 8.66, respectively, indicating that the mixed culture

could use this material as a growth substrate.

**Test condition** : A mixed microbial culture (protein concentration of 0.085 mg/l) was isolated

from an environment contaminated with organic cyanides and

polychlorinated biphenyls. This was grown for 48 hours on phosphate buffer (ph 7.0, 30 degrees C) containing propionitrile (1 g/l) as the sole source of carbon and nitrogen. The final concentration of protein, ammonia

and pH were determined.

**Test substance** : Test material was obtained from Aldrich Chemical Co. It is presumed that

the material has high purity.

**Reliability** : (4) not assignable

The study shows that the test material was used as a substrate (and therefore was metabolized); however, the extent to which the test material

biodegraded is difficult to determine from the study.

07.08.2003 (9)

#### 3.6 BOD5, COD OR BOD5/COD RATIO

#### 3.7 BIOACCUMULATION

#### 3.8 ADDITIONAL REMARKS

**Id** 107-12-0 4. Ecotoxicity Date 02.10.2003

#### 4.1 ACUTE/PROLONGED TOXICITY TO FISH

Type flow through

Species Pimephales promelas (Fish, fresh water)

Exposure period 96 hour(s) Unit mg/l **LC50** = 1520 EC50 = 1520 Limit test

Analytical monitoring

Method other: following the U.S. EPA Committee on Methods for Toxicity Tests

with Aquatic Organisms

Year **GLP** : no data

: as prescribed by 1.1 - 1.4 Test substance

Result : None of the controls or fish exposed to measured concentrations < = 887

mg/l died or had abnormal behavior. One fish exposed to 1100 mg/l died by 24 hours. All fish exposed to 2188 mg/l died by24 hours. Affected fish lost schooling behavior, were darkly colored and lost equilibrium prior to death. The 96 hour LC50 and EC50 values (with confidence intervals) were the

same (1520 and 1450-1580 mg/l, respectively).

The average (+/- SD) temperature, dissolved oxygen, hardness, alkalinity and pH of the water in the test chambers were 24.4 +/- 0.63 degrees C, 7.3 +/- 0.22 mg/l, 47.0 +/- 0.44 mg/l CaCO3, 40.1 +/- 1.04 mg/l CaCO3, and 7.6 +/- 0.21. It is not known whether these variables were affected by test material concentration.

Average and ranges of analytical concentrations of the chambers treated with 0, 455, 700, 1080, 1660 and 2550 mg/l material were <5, 375, 610, 885, 1098 and 2184 mg/l, respectively. When corrected for recovery (99.8 %), test material concentrations were < 5.01, 375, 611, 887, 1100 and 2188 mg/l.

The mean length and weight (+/- SD) of the fish at study termination were 20.8 +/- 1.673 mm and 0.092 +/- 0.0244 g.

: Newly hatched minnows from adults reared in flow-through tanks were held at 25 degrees C in flowing water with a 16-hr photoperiod and were fed brine shrimp nauplii three times daily (twice on weekends). They were cultured in filtered Lake Superior water or dechlorinated water from the city of Superior, WI (exact source not given) The two waters were similar in all measured chemical parameters. This water was used for test material dilution and all tests.

Healthy fish (32 days old) were fasted for 24 hours before treatment. They were pooled together in one tank and randomly distributed among the exposure chambers. Tests were initiated by adding 20 fish per treatment (455, 700, 1080, 1660 and 2550 mg/l) and control to test chambers containing 1.0 liter of water. Fish loading was 0.1278 g/l. The rate of exchange was 14.4 volumes of test water per day.

Observations of fish behavior and toxic signs were made at 2-8, 24, 48, 72 and 96 hr and recorded on checklists specifically formatted to convert observational data for approximately 100 endpoints into a numerically coded form. Death (cessation of opercular movements and inability to respond when prodded) was recorded at 24, 48, 72 and 96 hours. Dead fish were removed. At study termination, individual control fish were weighed (wet) and measured.

**Test condition** 

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> All test exposure chambers were sampled for test material concentration at the beginning of the test and daily thereafter. Concentrations of test material were analyzed using gas-liquid chromatography. All analyses included one spike and one duplicate sample for every 6 to 12 water samples.

> Five water quality parameters were routinely measured for each test: temperature, dissolved oxygen, total hardness, total alkalinity, and pH. The desired test temperature was 25 +/- 1 degrees C. Daily measurements of oxygen concentration were taken in each treatment and the control exposure chambers if fish were present. The control and one or more treatment chambers were sampled once for total hardness and alkalinity. The pH was measured once in the control and in one to five of the treatment tanks (specific times were not stated).

> The estimated LC50 and EC50 values, with corresponding 95% confidence intervals were calculated using the corrected average of the analyzed tank concentrations and the Trimmed Spearman-Karber Method. The EC50 values were based on loss of equilibrium manifested by an inability of the fish to remain in an upright position when swimming. The mean concentrations used in the calculations were corrected for analytical recoveries of spiked water samples.

: Purity of test material was 99 %. Test substance Reliability

: (1) valid without restriction.

The study was comparable to a guideline study.

Flag : Critical study for SIDS endpoint

10.08.2003 (20)

Type : static

Species Lepomis macrochirus (Fish, fresh water)

Exposure period 96 hour(s) Unit mg/l

**NOEC** < 10 measured/nominal LC50 = 41 measured/nominal

Limit test Analytical monitoring :

Method other: APHA, Standard Methods for Examination of Water and Wastewater,

14th Ed., 1975

: 1981 Year **GLP** : yes

Test substance : as prescribed by 1.1 - 1.4

Remark : Although the dissolved oxygen concentration in all 3 tested solutions

dropped below 40% saturation at 96 hours, the authors concluded that this

did not have an effect on mortality.

None of the negative control fish died. Ten percent of the fish exposed to Result

10 or 18 mg/l died by 96 hours. The mortality rate for fish exposed to 32 mg/l was 20% at 24 hours, 20% at 48 hours, 40% at 72 hours, and 50% at 96 hours. The mortality rate for fish exposed to 56 mg/l was 0% at 24 hours, 40% at 48 and 72 hours, and 50% at 96 hours. The mortality rate for fish exposed to 100 mg/l was 50% at 24 hours, and 90% at 48, 72 and 96 hours. The 24, 48 and 96 hour LC50 values for propionitrile (with confidence limits if applicable) were > 100 mg/l, 56 (43 - 78) mg/l and 41

(28 - 66) mg/l, respectively.

Negative control fish appeared normal at all observations. One fish in each of the 10 and 18 mg/l groups was surfacing at 96 hours. All fish that did not die after exposure to 32 mg/l appeared normal (with the exception of 1 fish that had an illegible observation at 48 hours). All survivors exposed to 56 and 100 mg/l were observed to be surfacing at 96 hours. The weight and

### 4. Ecotoxicity

ld 107-12-0 **Date** 02.10.2003

length of the fish at the end of the test were  $0.12 +/- 0.02 \, g$  and  $19 +/- 1.5 \, mm$ , respectively.

The temperature was 22 degrees C for all water samples. The dissolved oxygen concentration was 9.0 in each tested solution at 0 hours. Dissolved oxygen concentration ranged from 6.0 - 8.5 at 48 hours, and 2.0 - 4.3 at 96 hours. The water containing 100 mg/l propionitrile had the lowest dissolved oxygen concentration at 48 and 96 hours. The pH ranged from 7.1 - 7.7. Total ammonia concentrations were < 0.1 mg/l at each measurement. All temperatures, pH values and ammonia concentrations were within acceptable limits. Dissolved oxygen concentrations at 96 hours were not within acceptable limits (as defined by the protocol as 40-100% saturation).

The 24, 48 and 96 hour LC50 values for the positive control (with confidence limits if applicable) were > 0.00014 mg/l, 0.00012 mg/l and 0.00010 (0.000075 - 0.00014) mg/l, respectively. The LC50 values for the positive control were within the 95 % confidence limits reported in the literature.

**Test condition** 

Test organisms: The bluegill sunfish used in the study were obtained from Osage Catfisheries, Inc., Osage Beach, MO. All fish were on a 16 hour daylight photoperiod and observed for at least 14 days prior to testing. Fish received a standard commercial fish food daily until 48 hours prior to testing. The mean weight and length of the fish at the end of the test were 0.12 +/- 0.02 g and 19 +/- 1.5 mm, respectively. Maximum loading was 0.8 g fish/liter of solution.

Test material: Test concentrations were prepared based on the total compound. They were obtained by transferring appropriate aliquots from a working standard (150 mg/ml test material in absolute ethanol) directly to the test chambers. A preliminary, 48-hour range finding test was conducted with 1, 10 and 100 mg/l. The definitive, 96 hour test was conducted with 5 concentrations of test material ranging from 10 - 100 mg/l. The negative control chamber received an ethanol aliquot equivalent to the highest amount used in the test solutions. A positive control (Antimycin A), also was tested at concentrations ranging from 0.000014 to 0.00014 mg/l.

Test water: The well water from which the reconstituted water was prepared contained < 0.01 ppm aluminum, copper and zinc, <0.001 ppm arsenic, cadmium, and cobalt, 0.001 ppm chromium, 0.012 ppm iron, 0.009 ppm lead, <0.0001 ppm mercury, 0.0157 ppm nickel, and <0.3 ppb of commonly analyzed pesticides. The water was reconstituted to contain 48 mg/l NaHCO3, 30 mg/l CaSO4, 30 mg/l MgSO4, and 2 mg/l KCl. The hardness, alkalinity and initial pH of the water were 45 mg/l (as CaCO3), 35 mg/l (as CaCO3) and 7.3, respectively. The dissolved oxygen concentration at the start of the test was 9.0 mg/l. The temperature of the water was kept at 22 +/- 1 degrees C.

Test conduct: Tests were conducted in 5 gallon glass vessels containing 15 liters of reconstituted water. The test fish (10 per test concentration) were acclimated to the dilution water for 48 hours prior to testing. They were not fed during this acclimation period or during the test. The test concentrations (10, 18, 32, 56 and 100 mg/l) were chosen based on the results of a preliminary study. Two additional groups of 10 fish were exposed to the negative or positive control. Fish were added randomly within 30 minutes of preparation of the test solutions. All fish were observed at 24, 48, 72 and 96 hours for mortality and abnormal behavior. The pH, dissolved oxygen concentration, and temperature of water in the negative control, and 10 mg/l and 100 mg/l test vessels were determined at the beginning of the test and after 48 and 96 hours. Total ammonia concentration of water in these 3 vessels was determined at the beginning and end of the test.

Statistical analysis: Concentration vs. lethality data were analyzed by a computer program which utilized the binomial, moving average and probit tests to determine the LC50 values (and 95% confidence limits) at 24, 48 and 96 hours. The method of calculation selected for presentation (the probit method for propionitrile and the binomial method for Actinomycin A) was the one that gave the narrowest confidence limit.

**Test substance**: The purity of the test material (lot # 34) was 96.1%. Contaminants were not

mentioned.

**Reliability** : (2) valid with restrictions

Test concentrations were not analytically confirmed.

07.08.2003 (2)

Type : static

Species : Salmo gairdneri (Fish, estuary, fresh water)

Exposure period : 96 hour(s)
Unit : mg/l

NOEC : = 180 measured/nominal LC50 : = 340 measured/nominal

Limit test : no Analytical monitoring : no

Method : other: Committee on Methods for Toxicity Tests With Aquatic Organisms,

EPA-660/3-75-009, 1975

**Year** : 1981 **GLP** : yes

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: A brown film was present on the surface of water containing the highest 2

concentrations of propionitrile.

**Result**: None of the negative control fish or fish exposed to 100 or 180 mg/l died.

The mortality rate for fish exposed to 320 mg/l was 10% at 24 hours, 20% at 48 hours, 30% at 72 hours, and 40% at 96 hours. The mortality rate for fish exposed to 560 or 1000 mg/l was 100% by 24 hours. The 24, 48 and 96 hour LC50 values for propionitrile (with confidence limits if applicable) were 400 (320 - 560) mg/l, 380 (180 - 560) mg/l and 340 (180 - 560) mg/l,

respectively.

Negative control fish and fish exposed to 100 and 180 mg/l appeared normal at all observations. Surfacing was observed in some of the fish exposed to 320 mg/l at 48 hours (N = 8), 72 hours (N = 2) and 96 hours (N = 1). An illegible abnormality was observed in one fish exposed to 320 mg/l for 24 hours. The weight and length of the fish at the end of the test were 0.12 + - 0.02 g and 19 + - 1.5 mm, respectively.

The temperature was 12 degrees C for all water samples. The dissolved oxygen concentration was 9.4 and 8.7 in each tested solution at 0 and 48 hours, respectively. Dissolved oxygen concentration ranged from 8.0 - 9.1 at 96 hours. The pH ranged from 6.8 - 7.7 in all water samples tested (except for the water containing 1000 mg/l, which had a pH of 9.1 at 0 hours). Total ammonia concentrations were < 0.1 mg/l in the control and 100 mg/l solution at 0 hours, 0.21 mg/l in the control solution at 96 hours, 0.27 mg/l in the 1000 mg/l solution at 0 hours and the 100 mg/l solution at 96 hours, and 0.52 mg/l in the 320 mg/l solution at 96 hours. All temperatures, pH values, and dissolved oxygen and ammonia concentrations were within acceptable limits.

The 24, 48 and 96 hour LC50 values for the positive control (with confidence limits if applicable) were 0.00014 (0.00010 - 0.00047) mg/l, 0.000041 (0.000032 - 0.000052) mg/l and 0.000030 (0.000024 - 0.000042) mg/l, respectively. The LC50 values for the positive control were within the

95 % confidence limits reported in the literature.

**Test condition**: Test organisms: The rainbow trout used in the study were obtained from

Spring Creek Trout Hatchery in Lewistown, Montana. All fish were on a 16 hour daylight photoperiod and observed for at least 14 days prior to testing. Fish received a standard commercial fish food daily until 48 hours prior to testing. The mean weight and length of the negative control fish at the end of the test were 1.16 +/- 0.37 g and 44 +/- 3.7 mm, respectively. Maximum loading was 0.8 g fish/liter of solution.

Test material: Test concentrations were prepared based on the total compound. They were obtained by transferring appropriate aliquots from a working standard (150 mg/ml test material in absolute ethanol) directly to the test chambers. A preliminary, 48-hour range finding test was conducted with 10 and 100 mg/l. The definitive, 96 hour test was conducted with 5 concentrations of test material in a logarithmic series ranging from 100 - 1000 mg/l. The negative control chamber received an ethanol aliquot equivalent to the highest amount used in the test solutions. A positive control (Antimycin A), also was tested at concentrations ranging from 0.000014 to 0.00014 mg/l.

Test water: The well water from which the reconstituted water was prepared contained < 0.01 ppm aluminum, copper and zinc, <0.001 ppm arsenic, cadmium, and cobalt, 0.001 ppm chromium, 0.012 ppm iron, 0.009 ppm lead, <0.0001 ppm mercury, 0.0157 ppm nickel, and <0.3 ppb of commonly analyzed pesticides. The water was reconstituted to contain 48 mg/l NaHCO3, 30 mg/l CaSO4, 30 mg/l MgSO4, and 2 mg/l KCl. The hardness, alkalinity and initial pH of the water were 45 mg/l (as CaCO3), 35 mg/l (as CaCO3) and 7.3, respectively. The dissolved oxygen concentration at the start of the test was 9.4 mg/l. The temperature of the water was kept at 12 +/- 1 degrees C.

Test conduct: Tests were conducted in 5 gallon glass vessels containing 15 liters of reconstituted water. The test fish (10 per test concentration) were acclimated to the dilution water for 48 hours prior to testing. They were not fed during this acclimation period or during the test. The test concentrations (100, 180, 320, 560 and 1000 mg/l) were chosen based on the results of a preliminary study. Two additional groups of 10 fish were exposed to the negative or positive control. Fish were added randomly within 30 minutes of preparation of the test solutions. All fish were observed at 24, 48, 72 and 96 hours for mortality and abnormal behavior. The pH, dissolved oxygen concentration, temperature of water and total ammonia concentration in the negative control, and 100 mg/l and 1000 mg/l test vessels were determined at the beginning of the test. At 48 and 96 hours, these variables were tested in control water and water containing 100 and 320 mg/l test material (with the exception that total ammonia was not measured at 48 hours).

Statistical analysis: Concentration vs. lethality data were analyzed by a computer program which utilized the binomial, moving average and probit tests to determine the LC50 values (and 95% confidence limits) at 24, 48 and 96 hours. The method of calculation selected for presentation (the binomial method) was the one that gave the narrowest confidence limit.

**Test substance** 

: The purity of the test material (lot # 34) was 96.1%. Contaminants were not

mentioned.

**Reliability** : (2) valid with restrictions

Test concentrations were not analytically confirmed.

07.08.2003 (1)

#### 4.2 ACUTE TOXICITY TO AQUATIC INVERTEBRATES

Type : static

Species : Daphnia magna (Crustacea)

Exposure period : 48 hour(s)
Unit : mg/l

NOEC : = 100 measured/nominal LC50 : = 250 measured/nominal

Limit Test : no Analytical monitoring : no

Method : other: Committee on Methods for Toxicity Tests With Aquatic Organisms,

EPA-660/3-75009, 1975

**Year** : 1981 **GLP** : yes

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: A no observable effect level of 100 mg/l was listed by the authors although

abnormal behavior was noted in 1/20 daphnids exposed to this

concentration.

Result : None of the controls or daphnids exposed to 100 mg/l died during the

study. One daphnid in one vessel containing 180 mg/l died between 24 and 48 hours. Three daphnids exposed to this concentration in another vessel died (one by 24 hours). Therefore, the overall death rate of daphnids exposed to 180 mg/l was 5% and 24 hours and 20% at 48 hours. Ten out of 20 daphnids exposed to 320 mg/l (4 in one vessel and 6 in the other) died within 24 hours, and 15 died by 48 hours (8 in one vessel and 7 in the other). Therefore, the overall death rate of daphnids exposed to 320 mg/l was 50% and 24 hours and 75% at 48 hours. All daphnids exposed to 560 or 1000 mg/l died within 24 hours. The 24 and 48 hour LC50 values (with confidence limits) were 310 (270 - 350) mg/l and 250 (210 - 290) mg/l,

respectively.

In one vessel containing daphnids exposed to 100 mg/l, 1 daphnid was observed on the surface at 48 hours. Two daphnids exposed to 320 mg/l (in one vessel) were on the surface at 24 hours. Behavior of other

daphnids was normal.

The initial temperature, dissolved oxygen concentration and pH of the control water were 20 degrees C, 8.8 mg/l and 8.6. The temperature, dissolved oxygen concentration and pH of all water assayed at 48 hours were 21 degrees C, 7.8 - 8.3 mg/l and 8.6 - 8.7. All temperatures, dissolved

oxygen concentrations and pH values were within acceptable limits.

**Test condition** : Test organisms: The Daphnia magna used in the study were obtained from

an in-house culture. The adults were fed a suspension of trout chow and alfalfa daily until 24 hours prior to testing. All daphnids were held at 20 +/- 2 degrees C, under a 16 hour daylight photoperiod. First instar daphnids (< 24 hours old) were used in the test. Test daphnids were not fed during the

study.

Test material: Test concentrations were not corrected for sample purity. A primary standard of 20 mg/ml was prepared in water. Appropriate volumes of this standard were added to test water to obtain test concentrations.

Test water: The water used in the study was from a deep well source. It contained < 0.01 ppm aluminum, copper and zinc, <0.001 ppm arsenic, cadmium, and cobalt, 0.001 ppm chromium, 0.012 ppm iron, 0.009 ppm lead, <0.0001 ppm mercury, 0.0157 ppm nickel, and <0.3 ppb of commonly analyzed pesticides. The hardness, alkalinity, conductivity, dissolved oxygen concentration and initial pH of the well water were 255 ppm (as CaCO3), 368 ppm (as CaCO3), 50 micromhos/ cm, 9.2 ppm, and 7.8, respectively. The temperature of the water was kept at 20 +/- 1 degrees C.

Test conduct: Tests were conducted in 250 ml glass beakers containing 200 ml of well water. The test organisms (10 per test concentration) were added randomly to the test water within 30 minutes of addition of test material. The test concentrations (100, 180, 320, 560 and 1000 mg/l) were

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based on the results of a preliminary study performed with 1, 10 and 100 mg/l. One additional group of 10 organisms was exposed to water only served as a control. Each condition was tested in duplicate. All organisms were observed initially and after 24 and 48 hours of exposure for mortality and abnormal behavior (surfacing or loss of equilibrium). The pH, dissolved oxygen concentration and temperature of the control water were determined at the beginning and end of the study. Water containing 100, 320 and 1000 mg/l was analyzed for pH, dissolved oxygen concentration and temperature at the end (but not the beginning) of the study.

Statistical analysis: Concentration vs. lethality data were analyzed by a computer program which utilized the binomial, moving average and probit tests to determine the LC50 value (and 95% confidence limit) at 24 and 48 hours. The method of calculation selected for presentation (probit) was the

one that gave the narrowest confidence limit.

**Test substance** : The purity of the test material (lot # 34) was 96.1%. Contaminants were not

mentioned.

**Reliability** : (2) valid with restrictions

Test concentrations were not analytically confirmed.

Flag : Critical study for SIDS endpoint

01.08.2003 (3)

#### 4.3 TOXICITY TO AQUATIC PLANTS E.G. ALGAE

Species : Scenedesmus subspicatus (Algae)

Endpoint : biomass Exposure period : 96 hour(s) Unit : mg/l

**EC50** : = 789.303 calculated

Method : other Year : 2003 GLP : no

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: Measured inputs to the program are CAS No., melting point (-92.8 degrees

C), boiling point (97 degrees C), vapor pressure (39 mm Hg), and water

solubility (93,380 mg/l).

This is a supporting study for the SIDS endpoint.

**Reliability** : (2) valid with restrictions

15.08.2003 (14)

Species : Selenastrum capricornutum (Algae)
Endpoint : other: biomass and growth rate

Exposure period : 72 hour(s)
Unit : mg/l

NOEC : = 87.8 measured/nominal EC50 : > 87.8 measured/nominal

Limit test : yes Analytical monitoring : yes

Method : other: OECD: TG-201 and EEC/Annex V C.3

Year : 1999 GLP : yes Test substance : other TS

Remark : No protocol deviations were noted. The EbC50 (0-72 hr) and the ErC50 (0-

72 hr) were inestimable as greater than 50% inhibition in growth and/or biomass was not achieved. The significant loss (up to 80.7% over the course of the study) in test material was attributed to volatilization.

#### 4. Ecotoxicity

ld 107-12-0 **Date** 02.10.2003

#### Result

This is a supporting study for the SIDS endpoint for propionitrile.

Algae exposed to test material exhibited normal growth with respect to control. No deformed cells were noted. At the end of the test, the mean cell density in treated cultures was 1.365 x 10E6 cells /ml (compared to 1.356 x 10E6 cells in control).

The average concentrations of material in the test flasks at the beginning of the test and after 72 hours were 200.68 and 38.65 mg/l, respectively. Approximately 80.74% of the material was lost over the course of the experiment. The mean concentration was 87.82 mg/l. This concentration was listed as the NOEC.

Results of the photostability tests were similar to those in flasks containing test material and algae. The control solutions that were exposed to light or were in the dark exhibited 78.11% and 69.35% losses of test material.

The mean temperature and illumination were 24 degrees C and 746 foot-candles (range 744 - 748 foot-candles) throughout the test. The pH ranged from 7.42 - 7.88. The shaker speed was maintained at 100 rpm.

The test was considered to be valid since the mean cell concentration in control cultures increased by a factor of 136-fold within 72 hours.

: A 4-day culture of Selenastrum capricornutum SF-3148 (passage 5 in liquid algal medium) was used as the test algae. Several passages were performed prior to the test to confirm exponential growth.

Test medium: Sterile growth medium was prepared using high quality distilled water. The pH of the medium was measured and adjusted to 7.5 (+/- 0.1) using 0.01N NaOH.

Test material stock solution: Test material (0.156 ml) was added to 600.0 g of algal growth medium (to produce a nominal concentration of 200 mg/l). The solution was immediately capped and stirred for 1-2 minutes. An aliquot of the solution was removed for analysis of concentration at time 0.

Test conduct: All steps were carried out aseptically in a hood to prevent contamination. Test vessels were sterile 250 ml Erlenmeyer flasks. Test material stock solution (100 ml) was added to 5 flasks and test medium that did not contain test material was added to 3 flasks. Algae (515 microliters of algal stock culture to achieve an initial cell density of 1 x 10E4 cells/ml) were added to 3/5 flasks that contained test material and the three that did not. The two flasks that contained test material but were not inoculated served as photostability controls. One of the flasks was exposed to light and one was wrapped in foil to shield it from light. All flasks were secured with foam stoppers and transferred to a shaking incubator (24 degrees C, 100 rpm). They were illuminated at an average of 746.2 footcandles throughout the study.

Temperature, light intensity, and shaker speed (rpm) were assessed at the 0, 24, 48, and 72 hours. Concentrations of test material in the flasks that contained algae also were assessed at these times. The pH and was assessed at time 0 and after 72 hours. Concentrations of test material in the photostability controls also were measured at 0 and 72 hours. Concentrations of test material were analyzed using gas chromatography with flame ionization detection (GC/FID).

The exposure concentration was calculated as the geometric mean of the test concentrations analyzed at the 4 time points. Cell counts were performed after 24, 48 and 72 hours of exposure using a calibrated Coulter Counter. The mean algal cell count for the test and control curves was calculated. Two measures of growth [biomass (area under the growth

**Test condition** 

curve) and growth rate] were used to determine the effect of the material on algae. The concentrations that produced a 50% inhibition of biomass (EbC50) and growth rate (ErC50) relative to control were to be calculated by fitting linear regression models to the data.

The test was considered valid if the mean cell concentration in the control cultures increased by a factor of at least 16 within 72 hours.

The mean temperature and illumination were 24 degrees C and 746 foot-candles (range 744 - 748 foot-candles) throughout the test. The pH ranged from 7.42 - 7.88. The shaker speed was maintained at 100 rpm.

Test substance Conclusion

The test material was isobutyronitrile (CAS No. 78-82-0). Purity was 99.9%.
The results of this study indicate that the test substance would not be

classified according to the European Union's labeling directive and would correspond to a "low concern level" according to the U.S. EPA's

assessment criteria.

**Reliability** : (1) valid without restriction

This was a well-documented OECD-study conducted under GLP

assurances.

07.08.2003 (11)

Species : Selenastrum capricornutum (Algae)
Endpoint : other: biomass and growth rate

Exposure period : 72 hour(s)
Unit : mg/l

NOEC : = 133.4 measured/nominal EC50 : > 133.4 measured/nominal

Limit test : yes Analytical monitoring : yes

Result

Method : other: OECD: TG-201 and EEC/Annex V C.3

Year : 1999
GLP : yes
Test substance : other TS

Remark : Results of a pilot study conducted prior to this test indicated that a limit test

design would be appropriate for the material.

The EbC50 (0-72 hr) and the ErC50 (0-72 hr) were inestimable as greater than 50% inhibition in growth and/or biomass was not achieved. No

protocol deviations were noted.

This is a supporting study for the SIDS endpoint for propionitrile.

Algae exposed to test material exhibited normal growth with respect to control. At the end of the test, the mean cell density in treated cultures was

9.5 x 10E5 cells /ml (compared to 9.0 x 10E5 cells in control).

The average concentrations of material in the test flasks at the beginning of the test and after 72 hours were 206.0 and 85.7 mg/l, respectively. The mean concentration was 133.4 mg/l. This concentration was listed as the NOEC, EbC50 and ErC50.

Results of the photostability tests were similar to those in flasks containing test material and algae. The control solutions that were exposed to light or were in the dark exhibited 58% and 50% losses of test material.

The mean temperature and illumination were 24 degrees C and 747 foot-candles (+/- 5.5 foot-candles) throughout the test. The pH ranged from 7.4 - 7.6.

The test was considered to be valid since the mean cell concentration in control cultures increased by a factor of 90.2-fold within 72 hours.

**Test condition**: Test Organisms: A 4-day culture of Selenastrum capricornutum SF-3148

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(passage 3 in liquid algal medium) was used as the test algae. Several passages were performed prior to the test to confirm exponential growth. The density of cells in the stock culture was 2.58 x 10E6 cells/ml prior to use.

Test medium: Sterile growth medium was prepared using high quality distilled water. The pH of the medium was measured and adjusted to 7.5 (+/- 0.1) using 0.1N NaOH prior to use.

Test material stock solution: Approximately 0.151 ml (120 mg) of the test material was added to 600 ml of algal growth medium with a gas tight Hamilton syringe (to produce a nominal concentration of 200 mg/l). The solution was stirred for approximately 1 minute. An aliquot (1.0) of the solution was removed for analysis of concentration.

Test conduct: All steps were carried out aseptically in a hood to prevent contamination. Test vessels were sterile 250 ml Erlenmeyer flasks. Test material stock solution (100 ml) was added to 5 flasks and test medium that did not contain test material was added to 3 flasks. Algae (388 microliters of algal stock culture to achieve an initial cell density of 1 x 10E4 cells/ml) were added to 3/5 flasks that contained test material and the three that did not. The two flasks that contained test material but were not inoculated served as photostability controls. One of the flasks was exposed to light and one was wrapped in foil to shield it from light. All flasks were secured with foam stoppers and transferred to a shaking incubator (24 degrees C, 100 rpm). They were illuminated at 747 (+/- 5.5) footcandles throughout the study.

Temperature, light intensity, and shaker speed (rpm) were assessed at the 0, 24, 48, and 72 hours. Concentrations of test material in the flasks that contained algae also were assessed at these times. The pH and was assessed at time 0 and after 72 hours. Concentrations of test material in the photostability controls also were measured at 0 and 72 hours. Concentrations of test material were analyzed using gas chromatography with flame ionization detection (GC/FID). The exposure concentration was calculated as the geometric mean of the test concentrations analyzed at the 4 time points. Cell counts were performed after 24, 48 and 72 hours of exposure using a calibrated Coulter Counter. The mean algal cell count for the test and control curves. Two measures of growth [biomass (area under the growth curve) and growth rate] were used to determine the effect of the material on algae. The concentrations that produced a 50% inhibition of biomass (EbC50) and growth rate (ErC50) relative to control were to be calculated by fitting linear regression models to the data.

The test was considered valid if the mean cell concentration in the control cultures increased by a factor of at least 16 within 72 hours.

Test substance

The test material was butyronitrile (CAS No. 109-74-0). Purity was 99.9% (GC/FID).

Conclusion

The results of this study indicate that the test substance would not be classified according to the European Union's labeling directive and would correspond to a "low concern level" according to the U.S. EPA's assessment criteria.

(12)

Reliability

: (1) valid without restriction

This was a well-documented OECD-study conducted under GLP assurances.

07.08.2003

#### 4.4 TOXICITY TO MICROORGANISMS E.G. BACTERIA

# 4. Ecotoxicity **Date** 02.10.2003 4.5.1 CHRONIC TOXICITY TO FISH 4.5.2 CHRONIC TOXICITY TO AQUATIC INVERTEBRATES 4.6.1 TOXICITY TO SEDIMENT DWELLING ORGANISMS 4.6.2 TOXICITY TO TERRESTRIAL PLANTS 4.6.3 TOXICITY TO SOIL DWELLING ORGANISMS 4.6.4 TOX. TO OTHER NON MAMM. TERR. SPECIES **BIOLOGICAL EFFECTS MONITORING** 4.8 BIOTRANSFORMATION AND KINETICS 4.9 ADDITIONAL REMARKS

**Id** 107-12-0

#### 5.0 TOXICOKINETICS, METABOLISM AND DISTRIBUTION

In Vitro/in vivo : In vivo
Type : Excretion
Species : rat

**Number of animals** 

Males : 3

Females: 3

**Doses** 

Males : approximately 1.65 mg 14C-labeled priopionitrileFemales : approximately 1.65 mg 14C-labeled priopionitrile

Vehicle

Route of administration : gavage

Exposure time : Product type guidance : Decision on results on acute tox. tests :

Adverse effects on prolonged exposure :

Half-lives

1<sup>st</sup>: 2<sup>nd</sup>: 3<sup>rd</sup>:

Toxic behaviour Deg. product

Method : other
Year : 1987
GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: The overall 14C recovery was 92.05 %. The majority of the material was

eliminated in exhaled air or urine by 24 hours. An average of 27.05% of test material was detected in the Aquasol-2 bubbler (represents volatile organics) 0.5 hours after administration of 14C-labeleled propionitrile by gavage. By 3 hours, recovery in the 1N sodium hydroxide bubbler (material that had been exhaled as either 14CO2 or free cyanide) ranged from 38.54 to 49.25%. At 24 hours, the total 14C recovery in the urine (which represented a parent metabolite or thiocyanate) ranged from 0.76 to

5.83%. At 72 hours, a small percentage (< 2%) was found in the liver and

kidneys.

**Test substance**: Purity of the 14C-labeled propionitrile was 98.4%. The specific activity was

4.0 mCi/mmol.

**Conclusion**: The material is rapidly absorbed from the GI tract and eliminated through

expired air as parent material, free CO2 or free cyanide.

**Reliability** : (4) not assignable

The study was given a reliability rating of 4 because it was not reviewed in

detail.

10.08.2003 (7)

In Vitro/in vivo : In vivo
Type : Excretion
Species : rat

**Number of animals** 

Males: 3 Females: 3

**Doses** 

Males : 5.74 mg/kg, 7.9 microcuries Females : 7.35 mg/kg, 7.9 microcuries

Vehicle :

Route of administration : gavage

Exposure time

Product type guidance :

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Decision on results on acute tox. tests : Adverse effects on prolonged exposure :

Half-lives

1<sup>st</sup>: 2<sup>nd</sup>: 3<sup>rd</sup>:

Toxic behaviour

Deg. product

Method: otherYear: 1981GLP: no data

**Test substance**: as prescribed by 1.1 - 1.4

Remark : The total 14C-activity eliminated in 72 hours was 93.98 % and 74.73% of

the administered dose in males and females, respectively. After 72 hours, the 14C-activity found in the urine of 3 males and 3 females averaged 6.2% and 4.4% of dose, respectively. The cumulative 14C-activity after 72 hours found in the feces averaged 0.9% and 1.1% of the dose in males and females, respectively. The percentage of the dose recovered in whole blood cells was 0.38% and 0.31% for males and females, respectively. The majority of the 14C-activity (81.31% and 63.26% of the administered dose for males and females, respectively) was in respired air trapped in a sodium hydroxide bubbler (which represents CO2 or free cyanide). The amount of volatile material trapped in Aquasol-2 (which represented volatile organics) was 5.55% and 5.93% of dose for males and females,

respectively.

It was thought that the lower recovery in females was due to loss of 78 ml of NaOH from the bubbler for one rat. Assuming that this was not lost, the amount of material in the bubbler would have been 76.25 % of the dose. The total amount of 14C recovered in this animal would be 85.59%, which is in agreement with the average total amount of material recovered from

the other 2 females (84.3%).

**Test substance**: Purity of the 14C-labeled priopionitrile was 97.5%.

**Reliability** : (4) not assignable

The study is given a reliability rating of 4 because it was not reviewed in

detail.

05.08.2003 (8)

#### 5.1.1 ACUTE ORAL TOXICITY

Type : LD50

Value : = 40 mg/kg bw

Species : rat

Strain: Sprague-DawleySex: male/female

Number of animals : 60

Vehicle

**Doses** : 25.1, 31.6, 39.8, 50.1, 63.1 and 79.4 mg/kg bw

Method: otherYear: 1980GLP: no data

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: Whether the study was done according to GLP was not stated in the study

documents. However, an accompanying letter from the company that commissioned the study requested that the study should be performed

according to GLP. Therefore, it is assumed that it was.

**Result** : The numbers of animals that died after each of the doses is as follows:

Dose (mg/kg) Number of Deaths (all within 1 day)

25.1	1/5 males, 0/5 females
31.6	2/5 males, 1/5 females
39.8	4/5 males, 1/5 females
50.1	5/5 males, 2/5 females
63.1	3/5 males, 5/5 females
79.4	5/5 males, 4/5 females

The LD50 values [with 95% confidence limits (CL)] and slopes of the curves were as follows:

Sex	LD50 (mg/kg)	95 % CL (mg/kg)	slope
M/F M	40 38	35 - 46 28 - 51	4.9 3.5
F	52	43 - 63	5.6

Signs of toxicity included increasing weakness and collapse (both sexes), and tremors (in 2 males). Necropsies of animals that died revealed hemorrhagic lungs and liver, discoloration of liver, kidneys and spleen, and acute gastrointestinal inflammation. Average weights of survivors at days 7 and 14 were higher than initial weights. Viscera appeared normal in survivors.

#### **Test condition**

Thirty rats/sex were fasted overnight and divided into 6 groups of 5 rats/sex. These groups were given a single oral dose (undiluted) of propionitrile at 25.1, 31.6, 39.8, 50.1, 63.1 or 79.4 mg/kg. The average initial weights of males and females were 200 and 184 g, respectively. Animals were observed for 14 days, and weighed on days 7 and 14. It is presumed that they were observed daily. Animals that died were necropsied upon discovery. Survivors were euthanized and necropsied on day 14. The LD50 value, 95% confidence interval and slope of the curve were calculated by an unknown method.

Test substance

The approximate composition of the test material was 97% propionitrile, 1-

3% acrylonitrile, 1% adiponitrile and 2% water.

Reliability

: (2) valid with restrictions

The method of calculating the LC50 value was not listed.

13.08.2003 (38)

Type : LD50 Value :

Species : rat

Strain : Sprague-Dawley
Sex : male/female

Number of animals : 60

Vehicle :

**Doses** : 50.1, 63.1, 79.4, 100, 126, 158 mg/kg (males); 158, 200, 251, 316 mg/kg

(females)

Method : other Year : 1979 GLP : no data

**Test substance** : as prescribed by 1.1 - 1.4

**Result** : The numbers of animals that died after each of the doses is as follows:

Dose (mg/kg)	Number of Deaths (all in 1 - 3 days)
50.1 63.1 79.4 100	0/5 males 2/5 males 4/5 males 3/5 males
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126 3/5 males 158 4/5 males, 0/5 females 200 1/5 females 251 1/5 females 316 4/5 females

The LD50 values [with 95% confidence limits (CL)] and slopes of the curves were as follows:

Sex	LD50 (mg/kg)	95 % CL (mg/kg)	slope
М	75	58 - 98	3.9
F	270	235 - 310	9.0

Signs of toxicity in both sexes included weight loss (one to three days in survivors), increasing weakness, ocular discharge, tremors, convulsions, collapse and death. Diarrhea and dyspnea were also observed in females. Animals that died had hemorrhagic lungs, liver hyperemia and/or discoloration (in some instances), and gastrointestinal inflammation (acute

in some instances). Viscera appeared normal in survivors.

**Test condition**: Thirty males and 20 females were divided into 6 groups of 5 rats/sex.

Males were given a single oral dose (undiluted) of propionitrile at 50.1, 63.1, 79.4, 100, 126, or 158 mg/kg, and females were given 158, 200, 251 or 316 mg/kg. The average initial weights of males and 4 males were 237.5

and 242.5 g, respectively. Animals were observed for 14 days. It is presumed that they were observed daily. Animals that died were

necropsied upon discovery. Survivors were euthanized and necropsied on day 14. The LD50 value, 95% confidence interval and slope of the curve

for each sex were calculated by an unknown method.

**Test substance** : A data sheet containing information about the test material listed the purity

to be 90+%. Impurities included 0.05% acrylonitrile, < 0.1% water, and 0.16% adiponitrile. Additional impurities (other than an illegible one at 510

ppm) were not listed.

**Reliability** : (2) valid with restrictions

Test conditions are not described in detail.

10.08.2003 (39)

#### 5.1.2 ACUTE INHALATION TOXICITY

Type : LC50 Value : = 3.3 mg/l

Species : rat

Strain: Sprague-DawleySex: male/female

Number of animals : 60

Vehicle

**Doses** : 1.58, 2.51, 3.98, 6.31, 10.0, 15.8 mg/l

Exposure time : 4 hour(s)

Method : other

Year : 1978

GLP : no data

**Test substance** : as prescribed by 1.1 - 1.4

Remark : The NOAEL in ppm can be calculated using the following equation [ppm =

3300 mg/m3 x 24.45/55.09 (MW)]. For this experiment, the value is 1464.6.

**Result** : The numbers of animals that died after exposure to propionitrile is as

follows:

27 / 14

Concentration (mg/l)	Number of Deaths
----------------------	------------------

1.58	0/5 males, 0/5 females (0/10)
2.51	5/5 males 0/5 females (5/10)
3.98	5/5 males, 0/5 females (5/10)
6.31	5/5 males, 3/5 females (8/10)
10.0	5/5 males, 5/5 females (10/10)
15.8	5/5 males, 5/5 females (10/10)

All deaths occurred within 4 - 24 hours (with the exception of 1 male exposed to 3.98 mg/l, that died after 13 days).

The LD50 value for males and females combined [with 95% confidence limits (CL)] was 3.3 (2.6 - 4.3) mg/l. The slope of the curve was 4.1. These values were not calculated for males and females individually.

Signs of toxicity included salivation, lethargy, increasing weakness, tremors and convulsions, and collapse. Animals that died had hemorrhagic lungs, discolored liver (mottled in some) and acute gastrointestinal inflammation. Necropsies of survivors were normal.

**Test condition** 

**Test substance** 

Sixty rats (30/sex) were divided into 6 groups (5/sex/group). Each group was exposed to 1.58, 2.51, 3.98, 6.31, 10.0 or 15.8 mg/l by inhalation. During the test, males and females receiving the same concentration were placed together in 9 inch x 16 inch x 7 inch cages that were suspended in the middle of 210-liter drum-like chambers that were equipped with circulating fans. Test material was introduced into the chamber through a port equipped with a needle and syringe. No supplementary air was introduced. The average temperature and relative humidity inside the chambers were 24-25 degrees C and 70%, respectively.

Animals were observed for signs of toxicity during exposure and for 14 subsequent days. Animals that died were necropsied upon discovery. On day 14, survivors were euthanized and necropsied.

The LC50 value, 95% confidence limits and slope of the curve were calculated according to the method of DeBeer (reference was not given). The purity of the material was not listed. However, in a study conducted by

The purity of the material was not listed. However, in a study conducted by the same laboratory in 1979, a data sheet containing information about the test material listed the purity to be 90+%. Impurities included 0.05% acrylonitrile, < 0.1% water, and 0.16% adiponitrile. Additional impurities (other than an illegible one at 510 ppm) were not listed.

**Reliability** : (2) valid with restrictions

Basic data and methodologies are given.

Flag : Critical study for SIDS endpoint

10.08.2003 (40)

**Type** : LC100 **Value** : = 90.3 mg/l

Species : rat

Strain : Sprague-Dawley

Sex : male
Number of animals : 6
Vehicle :

 Doses
 : 90.3 mg/l

 Exposure time
 : 1.25 hour(s)

 Method
 : other

 Year
 : 1979

 GLP
 : no data

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: All six rats exposed to 90.3 mg/l for 1.25 hours died. The NOAEL in ppm

can be calculated using the following equation [ppm = 90300 mg/m3 x

24.45/55.09(MW)]. For this experiment, the value is 40076.87.

**Test substance** : A data sheet containing information about the test material listed the purity

to be 90+%. Impurities included 0.05% acrylonitrile, < 0.1% water, and 0.16% adiponitrile. Additional impurities (other than an illegible one at 510  $\,$ 

ppm) were not listed.

**Reliability** : (4) not assignable

The study was given a reliability rating of 4 because it was not reviewed in

detail.

10.08.2003 (39)

#### 5.1.3 ACUTE DERMAL TOXICITY

Type : LD50

Value : = 40 mg/kg bw

Species : rabbit

Strain : New Zealand white Sex : male/female

Number of animals : 40

Vehicle

**Doses** : 12.5, 25, 50 and 100 mg/kg **Method** : other: 40 CFR, Part 163.81-2

Year : 1981 GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

**Result**: The numbers of animals that died after each of the doses is as follows:

Dose (mg/kg) Number (and time) of Deaths

12.5 0/10 25 0/10

50 9/10 (4/5 M, 5/5 F), 3-23 hr

100 10/10, 2-7.5 hr

The LD50 values [with 95% confidence limits (CL)] were as follows:

Sex	LD50 (mg/kg)	95 % CL (mg/kg)
M/F	40	29 - 51
M	43	24 - 62
F	35	(could not be calculated)

Signs of toxicity included ataxia, convulsions, tremors, respiratory abnormalities (hyperpnea, hypopnea, dyspnea and arrythmic respiration), hypoactivity, prostration, hypothermia, nasal or ocular discharge, fecal staining, nasal discharge, soft stool, reddened nictitating membrane or reduced food consumption. Most animals in the 12.5 and 25 mg/kg dose groups were free of significant abnormalities from days 3 to 14.

Necropsies of animals that died revealed reddening of the nictitating membrane, mottling of the liver, GI abnormalities (reddened walls, black foci in the mucosa, black material adhered to the mucosa), and red interstitial walls or intestinal contents. Necropsies of survivors were normal.

Average weights of survivors at days 7 and 14 were generally higher than initial weights; however a few animals exhibited slight weight losses (0.1 or

0.2 kg). No dose-related differences in weight gain were apparent. Six and 3 of the animals given 12.5 and 25 mg/kg (respectively) had no dermal irritation. The other animals in these groups had only slight or very slight erythema without edema.

#### **Test condition**

Forty animals were acclimated for 38 days, then randomized to four groups of 5/sex. They were given food and water ad libitum. Animals considered unsuitable due to poor health or outlying weights were excluded. Pretest body weights were 2.6 - 3.3 kg for males and 2.5 - 3.6 kg for females. Approximately 18 hours prior to treatment, the hair on each rabbit was closely clipped from the trunk (dorsal and ventral surface and sides from scapular to pelvic area). Skin was not abraded. Test material (12.5, 25.0, 50.0 or 100 mg/kg) was applied directly onto the exposed skin and was spread evenly over the entire area. A layer of 8-ply gauze was then wrapped around the animal to cover the application site. This was wrapped in an impervious plastic sleeve, which was secured with masking tape. Elizabethan collars were placed on all animals.

Wrappings were removed after 24 hours, and the test site was wiped free of test material. Skin was scored for irritation according to the method of Draize 30 minutes after removal of the dressing. Animals were observed for toxicity at 1, 2, and 4 hours and daily thereafter for 14 days. Animals were weighed prior to treatment and 7 and 14 days after treatment. Animals that did not survive for 14 days were weighed and examined grossly at the time they were found dead. All animals surviving to day 14 were euthanized at this time and examined grossly.

The LD50 values (and 95% confidence intervals) for males, females and both sexes together were calculated using logarithmic-probit graph paper.

**Test substance** 

The purity of the test material was 94.5%. It also contained 1.1% adiponitrile, < 0.1% acrylonitrile, < 0.1% water, < 0.1 % solids and 0.07% para nitrosodiphenylamine.

**Reliability** : (1) valid without restriction

The test was performed according to an established guideline.

Documentation is thorough.

10.08.2003 (4)

Type : LD50

Value : = 56 mg/kg bw

Species : rabbit

Strain : New Zealand white Sex : male/female

Number of animals : 40 Vehicle :

**Doses** : 12.5, 25, 50 and 100 mg/kg **Method** : other: 40 CFR, Part 163.81-2

Year : 1981 GLP : no data

**Test substance**: as prescribed by 1.1 - 1.4

**Result**: The numbers of animals that died after each of the doses is as follows:

Dose (mg/kg) Number (and time) of Deaths

12.5 0/10 25 0/10

50 4/10 (3/5 M, 1/5 F), 3.5 - 21.5 hr 100 9/10 (5/5 M, 4/5 F), 4-21 hr

The LD50 values [with 95% confidence limits (CL)] were as follows:

LD50 (mg/kg)	95 % CL (mg/kg)
56 45	40 - 72 19 - 71
70	19 - 7 1 44 - 96
	56 45

Signs of toxicity included ataxia, convulsions, respiratory abnormalities (hypopnea, dyspnea and gasping), hypoactivity, prostration, nasal or ocular discharge, fecal staining or reduced food consumption. Necropsies of animals that died revealed reddening of the nictitating membrane, red or black foci or patches in the gastric mucosa, discoloration of liver or kidneys, red material in the GI tract or thickening of the abdominal wall. Necropsies of survivors were normal. Average weights of survivors at days 7 and 14 were higher than initial weights and were not affected by dose of test material. Approximately one half of the survivors at 24 hours had no dermal irritation. The others had only slight or very slight erythema without edema.

#### **Test condition**

Forty animals were acclimated for 35 days, then randomized to four groups of 5/sex. They were given food and water ad libitum. Animals considered unsuitable due to poor health or outlying weights were excluded. Pretest body weights were 2.7 - 3.2 kg for males and 2.6 - 3.4 kg for females. Approximately 18 hours prior to treatment, the hair on each rabbit was closely clipped from the trunk (dorsal and ventral surface and sides from scapular to pelvic area). Skin was not abraded. Test material (12.5, 25.0, 50.0 or 100 mg/kg) was applied directly onto the exposed skin and was spread evenly over the entire area. A layer of 8-ply gauze was then wrapped around the animal to cover the application site. This was wrapped in an impervious plastic sleeve, which was secured with masking tape. Elizabethan collars were placed on all animals.

Wrappings were removed after 24 hours, and the test site was wiped free of test material. Skin was scored for irritation according to the method of Draize 30 minutes after removal of the dressing. Animals were observed for toxicity at 1, 2, and 4 hours and daily thereafter for 14 days. Animals were weighed prior to treatment and 7 and 14 days after treatment. Animals that did not survive for 14 days were weighed and examined grossly at the time they were found dead. All animals surviving to day 14 were euthanized at this time and examined grossly.

The LD50 values (and 95% confidence intervals) for males, females and both sexes together were calculated using logarithmic-probit graph paper. The purity of the test material was not listed. It was used as received from

Monsanto. The density was listed as 0.7978 g/ml.

**Reliability**: (2) valid with restrictions

Purity of the material and documentation of GLP were not provided.

10.08.2003 (6)

Type : LD50

Value : = 90 mg/kg bw

Species : rabbit

Strain : New Zealand white Sex : male/female

Number of animals : 40

Vehicle

**Test substance** 

**Doses** : 12.5, 25, 50 and 100 mg/kg **Method** : other: 40 CFR, Part 163.81-21

Year : 1981 GLP : no data

**Id** 107-12-0 5. Toxicity Date 02.10.2003

Test substance

: as prescribed by 1.1 - 1.4

Result

: The numbers of animals that died after each of the doses is as follows:

Dose (mg/kg) Number (and time) of Deaths

12.5 0/10 25 0/10

50 1/10 (0/5 M, 1/5 F), 23.5 hr 6/10 (3/5 M, 3/5 F), 2.5-23.5 hr 100

The LD50 values [with 95% confidence limits (CL)] were as follows:

Sex	LD50 (mg/kg)	95 % CL (mg/kg)
M/F	90	65 - 115
M	90	49 - 131
F	85	35 - 135

Signs of toxicity included ataxia, convulsions, tremors, respiratory abnormalities (hyperpnea and dyspnea), hypoactivity, prostration, red eyes, nasal discharge, soft stool, fecal staining or reduced food consumption. Most animals in the 12.5 and 25 mg/kg dose groups and most survivors in the 50 and 100 mg/kg dose groups were free of significant abnormalities from days 2 to 14. Necropsies of animals that died revealed reddening of the nictitating membrane, pale liver, and GI abnormalities (reddened walls, black foci in the mucosa, and/or white film on the mucosa). Necropsies of survivors were normal.

Average weights of survivors at days 7 and 14 were higher than initial weights and were not affected by dose of test material. Approximately one half of the survivors at 24 hours had no dermal irritation. The others had only slight or very slight erythema, generally without edema.

**Test condition** 

Forty animals were acclimated for 31 days, then randomized to four groups of 5/sex. They were given food and water ad libitum. Animals considered unsuitable due to poor health or outlying weights were excluded. Pretest body weights were 2.5 - 3.1 kg for males and 2.7 - 3.4 kg for females. Approximately 25 hours prior to treatment, the hair on each rabbit was closely clipped from the trunk (dorsal and ventral surface and sides from scapular to pelvic area). Skin was not abraded. Test material (12.5, 25.0, 50.0 or 100 mg/kg) was applied directly onto the exposed skin and was spread evenly over the entire area. A layer of 8-ply gauze was then wrapped around the animal to cover the application site. This was wrapped in an impervious plastic sleeve, which was secured with masking tape. Elizabethan collars were placed on all animals.

Wrappings were removed after 24 hours, and the test site was wiped free of test material. Skin was scored for irritation according to the method of Draize 30 minutes after removal of the dressing. Animals were observed for toxicity at 1, 2, and 4 hours and thereafter for 14 days. Animals were weighed prior to treatment and 7 and 14 days after treatment. Animals that did not survive for 14 days were weighed and examined grossly at the time they were found dead. All animals surviving to day 14 were euthanized at this time and examined grossly.

The LD50 values (and 95% confidence intervals) for males, females and both sexes together were calculated using logarithmic-probit graph paper.

Test substance

The purity of the test material was not listed. It was used as received from Eastman Kodak. The density was listed as 0.7885 g/ml.

Reliability : (2) valid with restrictions

**Id** 107-12-0 5. Toxicity Date 02.10.2003

Purity of the material and documentation of GLP were not provided.

10.08.2003 (5)

#### 5.1.4 ACUTE TOXICITY, OTHER ROUTES

#### 5.2.1 SKIN IRRITATION

#### 5.2.2 EYE IRRITATION

#### 5.3 **SENSITIZATION**

#### REPEATED DOSE TOXICITY 5.4

Sub-chronic Type

Species rat

Sex male/female Strain Sprague-Dawley

: inhalation Route of admin.

: 14 weeks (total of 63 exposure days) Exposure period : 6 hours/day, 5 days per week

Frequency of treatm.

Post exposure period

Doses 60, 120, 209 ppm

Control group : yes

NOAEL < 60 ppm LOAEL = 60 ppmMethod : other Year 1984 **GLP** yes

Test substance as prescribed by 1.1 - 1.4

Result Test material concentrations: The nominal concentrations (+/- SD) were

56.0 +/- 8.8, 119.1 +/- 16.9 and 203.0 +/- 19.6. Corresponding analytical concentrations were 60.2 +/- 1.0, 120.3 +/- 1.1 and 209.0 +/- 1.3 ppm. Since the nominal analytical ratios were close to 1, the material was a true vapor. There was no indication that the test material was unstable. Distribution was uniform (> 96%) for each of the exposure concentrations. Airflow, temperature and relative humidity ranged from 1719-1765 l/min.

21.0 - 26.1 degrees C and 17-50%, respectively.

Effects at all exposure concentrations: Signs of toxicity (labored breathing, nasal discharge, salivation, discharge from the eyes, hypoactivity and/or alopecia) were observed in all exposed groups. Incidences of these signs increased in a dose-dependent manner. Males and/or females in all exposed groups had significant decreases in red blood cells and hemoglobin values. Urine thiocyanate concentrations increased in all exposed groups, with concentrations from animals exposed to 120 ppm similar to or higher than those exposed to 210 ppm. However, since a dose-dependent diuresis occurred, the total amount of urine thiocvanate present (concentration x urine volume) increased with increasing concentrations.

Effects at 209 ppm: Three males died or were killed in extremis (2 between exposures 2 and 3 and one between exposures 18 and 19). Arched back, ataxia, tremors or convulsions, biting, pawing or rubbing chin

against cage, irritation of the conjunctiva and breathing difficulties were noted in a few animals during exposure. Males and females exhibited significant decreases in body weight throughout the study (P <= 0.01). Average final body weights of males and females were lower than controls (377.5 g in exposed males vs. 435.4 g in controls and 255.0 g in exposed females vs. 276.6 g in controls). Absolute and/or relative heart, liver, spleen and kidney weights were increased in males and/or females. Absolute testes weights were decreased in males. Mean corpuscular hemoglobin concentrations (males only) were lower than control (all P <= 0.05). Serum alkaline phosphatase (males and females), SGOT (males only) and SGPT (males only) were increased, and BUN concentrations were decreased. No gross lesions were observed at termination. There was mildly increased hemosiderin deposition in the spleens of 10/15 females. Other microscopic changes were considered to be spontaneous.

Effects at 120 ppm: Ataxia was observed during exposure in 2 females. Males had significant body weight decreases (P <= 0.05) at two time points. Absolute and/or relative liver weights were increased in males and females and absolute and/or relative spleen weights were increased in males. Mean corpuscular hemoglobin concentrations (males only, P <= 0.05) were lower than control. No gross lesions were observed at termination. There was mildly increased hemosiderin deposition in the spleens of 11/15 females. Other microscopic changes were considered to be spontaneous.

Effects at 60 ppm: Absolute and relative spleen weights were increased in males. Serum thiocyanate concentrations were marginally increased in males and females.

Animals: Rats were acclimated for at least 10 days prior to use. Two days prior to the start of the study, males weighed 174-200 g and females weighed 132-145 g. On the first day of the study, the animals were 43 days old. Animals were randomly allocated by body weight into 4 groups of 15 animals/sex/group. Animals were individually housed in suspended mesh cages and given food and water ad libitum (except during exposure). Animal rooms were maintained at 70-74 degrees C and 35-60% relative humidity, with a 12 hour light/dark cycle.

Exposure conditions: Exposures (6 hr/day, 5 days/week) occurred in 10 m3 Rochester-style stainless steel and glass inhalation chambers. Rats were placed individually in wire mesh cages that were suspended in the chambers by 3-tiered racks. Males were placed on one side and females on the other. The concentrations of material in the chambers (20, 120 or 210 ppm) were controlled by either adjusting the nitrogen flow though the propionitrile in the bubblers or by changing the amount of test material in the bubblers. The bubbler was connected to a side port in the vertical particle-size separator, which was in turn connected to the air inlet at the top of the inhalation chamber. One bubbler was used in each of the generation systems. Airflow was maintained at a constant flow of 1727 liters/min. Nominal concentration measurements were determined daily for each chamber following exposure, by dividing the amount of test material delivered to the chamber (the difference between the pre- and postexposure weights) over the 6-hr exposure period by the total air volume during the same period. Concentrations of test material in the chambers were measured 4 times daily using a Miran 1A General Purpose Gas Analyzer. Additional samples of atmosphere from 9 specified locations in each chamber were also taken at 3 different times to determine if the vapor was distributed uniformly.

Test conduct: Animals were observed for clinical signs between the second and fifth hour of each exposure. Estimations of the percentages of animals exhibiting hypoactivity, eye irritation and breathing difficulties were made. All animals were individually examined for gross signs of toxicity preceding

**Test condition** 

and following each exposure and checked for mortality. Each animal was weighed and given a thorough examination for gross signs of toxicity on a weekly basis.

Animals were euthanized after 14 total weeks on the study. Terminal body weights were obtained (following an overnight fast). Blood and urine were collected. Whole blood was treated with an anticoagulant and was analyzed for total and differential erythrocyte count, total leukocyte count, platelet count, hematocrit, hemoglobin, and red blood cell indices (mean corpuscular volume, mean corpuscular hemoglobin, and mean corpuscular hemoglobin concentration). Serum was analyzed for albumin, globulin, total protein, blood urea nitrogen, total bilirubin, glucose, glutamic pyruvic transaminase (SGPT), alkaline phosphatase, glutamic oxaloacetic transaminase (SGOT), T3, T4, thiocyanate and lactate dehydrogenase. Urine was analyzed for the presence of thiocyanates.

Detailed necropsies were conducted on all rats that died during the course of the study, those that were killed moribund, and those that survived to study termination. The adrenal glands (both together), testes (with epididymides, heart, kidneys, liver, pituitary and spleen were weighed. The aforementioned organs and the following tissues were fixed in 10% neutral formalin: abdominal aorta, bone and bone marrow (femur), brain, esophagus, ovaries, colon, ileum, lung, lymph nodes (mesenteric), mammary gland, nasal turbinates, pancreas, thyroid/parathyroid, prostate, salivary gland, sciatic nerve, skeletal muscle, skin, spinal cord, stomach, thymus, trachea, urinary bladder, uterus (with cervix) and gross lesions. Eyes (with optic nerve) were fixed in a solution of 2% glutaraldehyde and 10% neutral buffered formalin. Tissues were processed, embedded in paraffin, cut at five microns, stained with hematoxylin and examined microscopically.

Statistical analyses: In life and terminal body weights and organ weight data were analyzed using Dunnett's test. Organ to body weight ratios were analyzed using the Mann-Whitney test, with the Bonferroni inequality. Data for frequencies of microscopic lesions were evaluated with the Fisher's exact test with the Bonferroni inequality. Hematological and serum and urine chemistry variables were examined using Dunnett's test.

Test substance Reliability : The purity of the test material was 96%. Impurities were not listed.

: (2) valid with restrictions

The study is comparable to a guideline study; however, a NOAEL was not

established.

Flag : Critical study for SIDS endpoint

10.08.2003 (34)

Type : Sub-acute

Species : rat

Sex : male/female
Strain : Sprague-Dawley

Route of admin. : inhalation Exposure period : 18 days

Frequency of treatm. : 6 hr/day, 5 days per week

Post exposure period : none

**Doses** : 44, 115, 329 ppm

Control group : yes

 NOAEL
 : = 115 ppm

 LOAEL
 : = 329 ppm

 Method
 : other

 Year
 : 1981

 GLP
 : no data

**Test substance**: as prescribed by 1.1 - 1.4

Result : Thirteen males and one female died. High dose animals exhibited

decreases in mean body weight, ruffled fur, irritability, tremors, and discharges around the nose and eyes. Hematological measurements were also depressed in high dose animals. No signs of toxicity or changes in hematological measurements were observed in animals exposed to 44 or 115 ppm. There was no effect of treatment on clinical chemistries or urinalyses. No treatment-related gross or microscopic lesions were

observed.

**Test condition**: Groups of 18 animals/sex were exposed to 0 (control) 44, 115 or 329 ppm,

6 hr/day for 18 days.

**Test substance**: Purity of the test material was 94.87%.

**Reliability** : (4) not assignable

The study was given a reliability rating of 4 because it was not reviewed in

detail.

10.08.2003 (29)

#### 5.5 GENETIC TOXICITY 'IN VITRO'

Type : Mammalian cell gene mutation assay
System of testing : L5178Y Mouse Lymphoma Cells
Test concentration : 2143 to 5000 micrograms/ml
Cytotoxic concentr. : > 5000 micrograms/ml

Metabolic activation: withoutResult: negativeMethod: otherYear: 1982GLP: ves

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: An initial toxicity test showed that complete toxicity was not reached at

5000 micrograms/ml.

**Result** : First study: After the 2 day expression period, ten cultures (2 per

concentration) were cloned at 2714, 3286, 3857, 4429, and 5000 micrograms/ml. All cultures exhibited mutant frequencies similar to controls (ranged from 0.2 - 0.4 per 10E4 surviving cells) and the total

growth ranged from 61 to 106%.

Second study: After the 2 day expression period, twelve cultures (2 per concentration) were cloned at 2143, 2714, 3286, 3857, 4429, and 5000 micrograms/ml. All cultures exhibited mutant frequencies similar to controls (ranged from 0.2 - 0.3 per 10E4 surviving cells) and the total growth ranged from 69 to 94%.

Based on the combined results of both assays, none of the test concentrations induced a positive response (all p values were > 0.01). The linear component of the dose-response curve also was not statistically significant.

The test was considered valid, as the control frequencies were within the required range (0.2 - 0.3 per 10E4 surviving cells in the two studies) and EMS induced 9.2 and 7.6 mutants per 10E4 surviving cells at 0.5 microliters/ml and 28.4 and 24.5 mutants per 10E4 surviving cells at 0.5 microliters/ml and 1.0 microliters/ml, in the 2 studies, respectively.

**Test condition** 

Cell preparation: Prior to use in the test, L5178Y cells that were actively growing in culture were cleansed as described by Clive et al. (Mutat Res 31:17-29, 1975). Three ml of thymidine, hypoxanthine, methotrexate and glycine (THMG) stock solution was added to 100 ml cell suspension containing 0.1 x 10E6 cells/ml. The culture was gassed with 5% CO2 in air and placed in a shaking incubator (125 rpm, 37 degrees C). After 24 hours, the THMG was removed by pelletizing the cells and decanting off the supernatant. The cells were rinsed in 20 ml of F10 P (Fisher's Media for

leukemic cells with 10% heat inactivated horse serum) and reinstated in culture at 3 x 10E 4 cells/ml in 100 ml of F10 P plus 1 ml of THMG stock solution.

Test conduct: The test material was solubilized and diluted to produce dose levels from 2714 to 5000 micrograms/ml. Each concentration was tested in duplicate, and the test was performed twice. An additional concentration of 2143 micrograms/ml was tested in one of the experiments. All stock solutions were freshly prepared. Four ml of FoP was added to each tube. This yielded a final cell suspension of 0.6 x 10E6 cells/ml. Two control tubes received solvent only (DMSO). Positive controls were treated with ethyl methane sulfate (1.0 and 0.5 microliters/ml). All tubes were gassed with 5% CO2 in air and placed on a roller drum apparatus for 4 hours at 37 degrees C. All steps were carried out under amber lighting and the cells were incubated in the dark for 4 hours. After 4 hours, the cells were washed twice with 10 ml of F10P, then resuspended in 20 ml of F10P gassed with 5% CO2 in air, and replaced on a roller drum apparatus at 37 degrees C.

After the initial exposure, the cells were incubated for 2 days with a cell population adjustment (to 0.3 x 10E6 cells/ml) at 24 and 48 hours. At the end of this expression period, cells (a 2 x 10E-2 dilution of 1.5 x 10E6 cells/ml) were placed in 100 ml cloning medium containing 0.37% noble agar, and incubated in a shaking incubator at 37 degrees C. Trifluorothymidine (TFT; final concentration of 3 micrograms/ml) was added to one of the duplicate flasks per concentration. After 15 minutes, the flasks were removed, and 33 ml of the cell suspension from each flask was pipetted into 3 100 mm Petri plates. To accelerate the gelling process, the plates were stored at 4 degrees C for 20 min. The plates were then removed and incubated at 37 degrees C in a humidified 5% CO2 atmosphere for 10-12 days.

After the incubation period, the plates were scored for total number of colonies per plate. The plates that did not contain TFT served as viability controls. Each plate was counted 3 times by an automatic colony counter, and the median count was recorded. The mutation frequency was determined by dividing the average number of colonies in the 3 TFT plates by the average number of colonies x 10E4 in the 3 viability control plates and multiplying the quotient by 2.

Validity criteria: The test was considered acceptable if the positive control induced at least a 2-fold increase in the frequency of mutants with respect to the solvent control and resulted in a viability rate of 10-80%. The solvent control frequency also had to be 0.2 - 1 per 10E 4 cells, the plating efficiency of the control at least 50%, and the material tested to either 500 micrograms/ml, or at the limit of 10% viability or solubility. A test was considered positive if at least 2 concentrations that caused no more than 90% toxicity caused 2-fold increases in the frequency of mutants with respect to solvent controls and the response was concentration-dependent (p < 0.01) for at least 2 concentrations that did not cause > 90% cytotoxicity. A test was considered negative if none of the concentrations caused a significant increase in the frequency of mutations (p > 0.01) and the linear component of the dose-response curve was not significant (p > 0.1) for test concentrations resulting in at least 10% relative total growth.

**Test substance** 

The test material was propionitrile, used as received from Kodak. The purity was listed as 97.8%. Impurities included adiponitrile (0.3%), paranitrosophenylamine (0.1%), water (0.1%), acrylonitrile (<0.1%), and solids (<0.1%).

Reliability

: (1) valid without restriction

The study is comparable to a guideline study.

Flag 07.08.2003 : Critical study for SIDS endpoint

(27)

**Id** 107-12-0 5. Toxicity Date 02.10.2003

Type : Ames test

System of testing S. typhimurium strains TA98, TA100, TA1535 and TA1538

Test concentration up to 10,000 micrograms per plate Cytotoxic concentr. > 10,000 micrograms per plate

Metabolic activation with and without

negative Result Method other 1977 Year **GLP** no

Test substance as prescribed by 1.1 - 1.4

: The authors stated that the test was negative, even though there was a Remark

positive response with metabolic activation at one concentration in one

Result Experiment 1: There was no effect of treatment with test material on the

number of revertants in any strain in the absence or presence of S-9. The numbers of revertants per plate in control strains TA98, TA100, TA1535 and TA1538 without microsomes were 13, 42, 11 and 11, respectively. The numbers of revertants in strains TA98, TA100, TA1535 and TA1538 treated with 5 to 1000 micrograms/plate test material without microsomes ranged from 3-12, 24-47, 6-19 and 5-9, respectively. The numbers of revertants per plate in control strains TA98, TA100, TA1535 and TA1538 with microsomes were 47, 47, 11 and 40, respectively. The numbers of revertants in strains TA98, TA100, TA1535 and TA1538 treated with 5 to 1000 micrograms/plate test material with microsomes ranged from 33-50. 24-44, 7-13 and 12-39, respectively. The positive controls induced 11, 104 and 26 revertants per plate in strains TA98, TA100 and TA1538 without microsomes and 358, 832 and 202 revertants/plate with microsomes.

Experiment 2: There was no effect of treatment with test material on the number of revertants in any strain in the absence of S-9. The numbers of revertants per plate in control strains TA98, TA100, TA1535 and TA1538 without microsomes were 7, 37, 7 and 8, respectively. The numbers of revertants in strains TA98, TA100, TA1535 and TA1538 treated with 2500 to 10000 micrograms/plate test material without microsomes ranged from 4-7, 3-36, 7-12 and 4-11, respectively. The numbers of revertants per plate in control strains TA98, TA100, TA1535 and TA1538 with microsomes were 36, 34, 5 and 26, respectively. The numbers of revertants in strains TA98, TA100, and TA1538 treated with 2500 to 10000 micrograms/plate test material with microsomes ranged from 28-41, 25-31, and 19-31, respectively. In strain 1535 in the presence of microsomes, 4, 555, 7 and 9 revertants/plate were found after incubation with 2500, 500, 7500 and 10000 micrograms/plate. The response at 5000 micrograms/plate appeared to be an aberration, since it was not dose-dependent. The positive controls induced 15, 1011 and 20 revertants per plate in strains TA98, TA100 and TA1538 without microsomes and 331, 1395 and 299 revertants/plate with microsomes.

The enzyme controls and histidine response check all gave predicted responses.

**Test condition** Two plate tests were performed - one with 5, 10, 25, 50, 100, 500 and 1000

> micrograms/plate and another with 2500, 5000, 7500 and 10000 micrograms/plate. The vehicle for the test material was water. Both tests were conducted in the presence and absence of liver microsome preparations from Aroclor-induced rats (sex not stated). The positive controls for strains TA98, TA100, TA1535 and TA1538 were

benzo(a)pyrene. 2-aminoanthracene, diethyl sulfate, and 2-

aminoanthracene, respectively (doses were not stated). Enzyme controls and histidine response checks also were performed.

Reliability : (4) not assignable

Purity of the test material was not given. Only 4 strains were tested. Numerical results for the positive control in strain TA1535 were not given (only listed as a positive sign). Documentation is limited. It appears that the assay was not performed in triplicate. The positive control for TA98 without microsomes did not induce an increase in revertants in the first experiment. Criteria for a positive test were not mentioned. The test that gave a positive result at one concentration should have been repeated.

10.08.2003 (19)

Type : Unscheduled DNA synthesis
System of testing : Primary rat liver cell cultures
Test concentration : up to 2500 micrograms/ml
Cytotoxic concentr. : 5000 micrograms/ml

Metabolic activation

Result : negative
Method : other
Year : 1985
GLP : yes

**Test substance** : as prescribed by 1.1 - 1.4

Remark : The hepatocytes would be expected to metabolize the material, due to the

presence of cytochrome p450 and other enzymes in the cells.

A positive control (2-acetylaminofluorene) which is known to induce unscheduled DNA synthesis after metabolic conversion to its active form

was positive in the test.

**Test substance**: Purity of the test material was 97 +/- 1%.

**Reliability** : (4) not assignable

The study is given a reliability rating of 4 because it was not reviewed in

detail.

07.08.2003 (32)

#### 5.6 GENETIC TOXICITY 'IN VIVO'

Type : Cytogenetic assay

Species : rat

Sex: male/femaleStrain: Sprague-Dawley

Route of admin. : gavage

Exposure period : 6, 24 or 48 hours
Doses : 0, 100 and 200 mg/kg

Result : negative
Method : other
Year : 1985
GLP : yes

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: The doses used in this study were based on a preliminary study performed

with 30, 100, 300, 500 and 750 mg/kg, which showed abnormal clinical signs and loss of body weight at doses > = 100 mg/kg, and no effect on mitotic index at 100 or 300 mg/kg. Initially, 200 mg/kg was chosen as the dose for both males and females. However, since 9/15 males administered 200 mg/kg died within 24 hours, the dose in males was reduced to 100

mg/kg.

Result : The number of metaphases analyzed in control animals at each time point

ranged from 475-500 cells (from a total of 10 animals). Two hundred and fifty metaphases from 5 animals were analyzed at each time point for animals treated with propionitrile (with the exception of 100 cells isolated at 48 hours from 2 females treated with 200 mg/kg). There was no effect of

treatment with propionitrile on the frequency of chromosomal aberrations (0 - 0.4 % aberrant cells per group in treated vs. 0 - 0.2 % in controls and 0 - 0.004 aberrations per cell in treated vs. 0 - 0.002 in control), mean chromosome number (treated ranged from 41.74 - 41.84 and control ranged from 41.75 - 41.83) or mitotic index (treated ranged from 0.52 - 2.40 and control ranged from 1.20 - 2.48) at any time point. A significant increase in the percentage of aberrant cells (25.52 in treated vs. 0 in control) and average number of aberrations per cell (1.401 in treated vs. 0 in control), and a decrease in mitotic index (0.28 vs. 1.20 in control) was observed in cells from animals treated with cyclophosphamide (the positive control).

Test condition

Three females treated with 200 mg/kg died. All males and females treated with propionitrile exhibited signs of toxicity. Males exhibited depression, red stains on nose/eyes, soft feces, dilated pupils and urine stains, and females exhibited depression, cold to touch, red stains on nose/eyes, wheezing, urine stains and tremors. Reduced body weights were observed in treated males at 24 hours and females at 24 and 48 hours. Eighty one Sprague-Dawley rats/sex (approximately 46-51 days old) were acclimated for 19 days prior to treatment. Food and water was supplied ad libitum. Eighty animals/sex were randomized and 35 per sex were assigned to the study. A single dose of test material (10 ml/kg, corrected for 100% purity) was administered by oral gavage to 2 groups of 15 male rats each at 0 (corn oil vehicle control) or 100 mg/kg and 2 groups of 15 females at 0 (vehicle control) or 200 mg/kg. Five animals per sex were euthanized at approximately 6, 24 and 48 hours after administration of test material. An additional 5 animals per sex were given the positive control cyclophosphamide (40 mg/kg) and euthanized 24 hours after treatment.

Animals were observed twice daily for general appearance, behavior, and clinical signs. Body weights of all animals were recorded just prior to administration of test material and just prior to colchicine administration (for animals that were to be killed 24 and 48 hours after treatment).

Approximately 4, 22 and 46 hours after test material was given, the appropriate groups of animals received a single intraperitoneal injection of colchicine (2.0 mg/kg body weight, dosing factor 5 ml/kg) to inhibit mitosis and arrest cells in metaphase. The colchicine was dissolved in Hank's Balanced Salt Solution (HBSS). Animals were euthanized approximately 2 hours after colchicine injection.

Bone marrow cells were collected from both femurs of each animal by aspiration into 5 ml of HBSS heated to 37 degrees C. The aspirate was spun in a centrifuge for 5 minutes at approximately 1100 rpm. The supernatant was decanted and 5.0 ml of preheated 0.075 M KCl was added to each tube. After 25 minutes, five drops of freshly prepared fixative (methanol:acetic acid, 3:1) were added to each tube. The tubes were capped, inverted and spun in a centrifuge for 5 minutes at 1100 rpm. The cells were resuspended in 5 ml of fixative, and again spun in a centrifuge for 5 min at 1100 rpm. This procedure was repeated two more times, and the cells were suspended in fresh fixative and refrigerated. After chilling, the cells were spun in a centrifuge at 1100 rpm for 5 minutes, the supernatant was decanted, and the cells were resuspended in 0.5 - 2 ml fresh fixative. Several drops of the final cell suspension were dispersed onto microscope slides and air-dried. Two to four slides were made per animal and were marked with the animal's identification number.

The cells on the slides were stained in fresh Giemsa for 10 minutes, rinsed twice with distilled water, air-dried, and mounted with glass coverslips. Code numbers were then blindly assigned to the slides. The slides were

not decoded until all had been analyzed.

At least 50 cells in metaphase were analyzed per animal (if possible). Otherwise, as many spreads as possible were analyzed. The slides were scanned with a low power objective (10 or 25 X) and the chromosomes were analyzed with a high power oil immersion lens (100X). Only those cells in metaphase were analyzed for cytogenetic abnormalities. The following items were recorded for each animal: numbers and types of chromosome aberrations (chromatid and chromosome breaks, chromatid and chromosome gaps, exchanges, cells with >= 10 aberrations, and pulverized cells), mitotic index, chromosome number for each metaphase and the vernier location of each metaphase containing damage.

The mean mitotic indices, chromosome numbers, percent aberrant cells and the mean number of aberrations per cell for each group were statistically compared using the Kruskal-Wallis nonparametric analysis of variance and nonparametric pairwise group comparisons. Body weight data were analyzed by the analysis of covariance. All tests were evaluated at the one-tailed 95% confidence interval (p < 0.05).

**Test substance** : Purity of the test material was 95.7 %. Impurities included 3.0%

acrylonitrile, 0.9% adiponitrile, and 0.1% solids (not identified).

**Reliability** : (2) valid with restrictions

Only two animals were available for analysis at the highest dose tested

(200 mg/kg).

07.08.2003 (21)

#### 5.7 CARCINOGENICITY

#### 5.8.1 TOXICITY TO FERTILITY

Type : Fertility
Species : rat
Sex : female

Strain : Sprague-Dawley Route of admin. : inhalation

**Exposure period** : 21 to 33 days (depending on day of mating)

Frequency of treatm. : 6 hr/day, 7 days/week

Premating exposure period

Male : 0 days Female : 21 days

**Duration of test** : to gestation days 13-15

No. of generation :

studies

**Doses** : 60, 120 and 210 ppm

Control group : yes NOAEL parental : = 60 ppm other: NOAEL : = 210 ppm

**Reproductive Toxicity** 

Method : other Year : 1984 GLP : yes

**Test substance** : as prescribed by 1.1 - 1.4

Result : Exposure concentrations: The average mean daily analytical exposure

concentrations (60.1, 120.2 and 209.2 ppm) were within 1% of the target levels (60, 120 and 210 ppm, respectively). Mean temperatures and humidities ranged from 22-25 degrees C and 26-29%, respectively.

Signs of toxicity: None of the animals died. There was no effect of test material on body weight. Animals exposed to 210 ppm exhibited arched back (N = 4 on days 1-10 and N=2 on days 11-20), lacrimation (N = 2 on days 1-10 and N = 1 on days 21-30), salivation (N= 15 on days 1-10, N = 22 on days 11-20 and N = 21 on days 21-30) hypoactivity (N = 13 on days 1-10, N = 5 on days 11-20 and N = 3 on days 21-30), staining of facial fur (N = 2 on days 1-10, N = 4 on days 11-20 and N = 4 on days 21-30) and red nasal encrustation (N = 1 on days 1-10, N = 5 on days 11-20 and N = 5 on days 21-20) after exposure. Animals exposed to 120 ppm also exhibited salivation (N = 6 on days 11-20 and N = 4 on days 21-30, staining of facial fur (N = 7 on days 1-10, N = 5 on days 11-10 and N = 2 on days 21-30)and red nasal encrustation (N = 2 on days 1 1-0, N = 8 on days 11-20 and N = 6 on days 21-30). A few animals in the 60 ppm group also exhibited red nasal encrustation (N = 1 on days 1-10, N = 3 on days 11-20 and N = 1 on days 21-30) and staining of facial fur (N = 1 on days 1-10, N = 3 on days 11-20 and N = 1 on days 21-20). One control animal had stained facial fur on days 21-30 and another had red nasal encrustation on days 1-10. Signs of toxicity generally abated by the morning after exposure. Alopecia was observed in animals (N = 2 controls, N = 3 low dose, N = 5 mid dose, N = 9) at one or more of their weekly physical examinations.

The only remarkable findings at gross necropsy were bilateral uterine hydrometra in one animal exposed to 210 ppm and hydrometra in the left uterine horn of one animal exposed to 120 ppm.

Fertility: There was no effect of treatment on fertility. Efficiency of mating (32.0%, 32.0%, 30.7% and 25.0% in the control, low, mid and high dose groups) and pregnancy rate (100%, 95.8%, 100% and 91.3% in the respective groups) were comparable between groups. There was no difference in the numbers of live implants (ranged from 13.4 - 13.9), resorptions (ranged from 0.6 - 0.8), nidations (ranged from 14.1 - 14.5), corpora lutea (ranged from 13.0 - 15.2), preimplantation loss (4-8%) and postimplantation loss (4-6%). Evaluation of the vaginal smears of 2 females that did not copulate showed one that did not cycle (but was pregnant at necropsy), and another that only went through the cycling stage of proestrus.

**Test condition** 

Animals: Virgin female Sprague Dawley rats (43 days old upon receipt) and virgin male Sprague Dawley rats (50 days old upon receipt) were acclimated for one week and examined for general health and the presence of pinworms and ectoparasites. Body weights of ten females and ten males that were taken upon receipt were 128-144 g and 178-233 g, respectively. No significant health problems were noted during the acclimation period, and the animals were released for study. Males and females were individually caged (except during mating). Food and water were available ad libitum (except food was not available to females during exposure). Animals were housed in a room maintained at 72 +/- 2 degrees F and 40-60% relative humidity, under a 12 hr light/dark cycle.

Exposures: Exposures (6 hr/day, 7 days/week) occurred in 10 m3 Rochester-style stainless steel and glass inhalation chambers. Due to inclement weather and building equipment failures, 2 exposures (days 2 and 16) were only for 4 hours and one exposure (day 1) was for 5 hours. Atmospheres of propionitrile vapor were generated using bubbler systems. The concentrations of material in the chambers were controlled by either adjusting the nitrogen flow though the propionitrile in the bubblers or by changing the amount of test material in the bubblers. Concentrations of test material in the chambers were measured 4 times daily (except during day 16) using a Miran 1A General Purpose Gas Analyzer. The nominal concentration of material, temperature and humidity also were measured (at intervals that were not listed).

Study conduct: Twenty four females per group were assigned to be

**Id** 107-12-0 5. Toxicity Date 02.10.2003

> exposed to target concentrations of 0, 60, 120 or 210 ppm test material. Exposure began when females were 63 days old. Animals were observed during exposure for signs of toxicity. After 21 days of exposure (which was sufficient to cover 3-4 estrus cycles), females were randomly mated (1:1) to an untreated male that had been assigned to the corresponding treatment group (30 males were assigned per group). At night, after exposure. females were caged with their assigned male until copulation was confirmed (by presence of a copulatory plug or sperm in vaginal smear) or 5 nights without confirmed copulation. Females that failed to mate with the assigned male were mated with another male that had copulated with another female in the same group. Nightly co-housing with the second male occurred until copulation was confirmed (or for a maximum of 7 nights). The day on which copulation was confirmed was considered gestation day 0. Exposure of females continued until copulation was confirmed or a maximum of 12 nights of cohabitation with males without signs of copulation. Vaginal smears were taken on 5 consecutive days for females that did not exhibit copulation.

After assignment to the study, males and females were weighed once per week. Mated females were weighed on gestation days 0 and 13. Females were given a thorough physical examination once per week and were observed for clinical signs of toxicity before and after exposures. All animals were checked twice daily for mortality and gross abnormalities.

Females were killed on gestation day 13 (or the nearest working day after gestation day 13, up to gestation day 15). Females without confirmed copulation ware euthanized in the second week after the last day of cohousing. Each female was given an external examination and weighed. The tissues and organs of the thoracic and abdominal cavities were examined for gross lesions. Pregnancy status was determined, nidation sites and were classified, and corpora lutea were counted. The ovaries and uteri of females were preserved in 10% neutral buffered formalin. Males were killed after mating and were not examined.

Statistical analyses: Body weight data were analyzed using Dunnett's test. Mating and pregnancy rate data were analyzed by a Fisher's exact test and an uncorrected chi-square test. Other data were analyzed by the Mann-Whitney U test. The Bonferroni inequality was assumed when comparing multiple treatments to control values for all tests except Dunnett's test. The critical level for significance was p < 0.05.

Purity of the test material was 96.1%. Impurities included acrylonitrile **Test substance** 

(0.1%), adiponitrile (0.7%), p-nitrosodiphenylamine (0.08%) and water (< 0.1%). Analyses indicated no significant decomposition of the test material

over the course of the study. The authors concluded that the incidences of red nasal encrustation in the

low dose animals, alopecia in the mid and high dose animals and staining of facial fur in all treated groups were too low to be definitely related to administration of test material. There was no effect of treatment on fertility

of females.

Reliability (1) valid without restriction

Study is comparable to a guideline study.

: Critical study for SIDS endpoint Flag

07.08.2003 (24)

Fertility **Type Species** rat Sex male

Strain Sprague-Dawley Route of admin. inhalation

46 to 57 days (depending on day of mating) Exposure period

Frequency of treatm. 6 hours/day, 5 days/week

Premating exposure period

Conclusion

**Id** 107-12-0 5. Toxicity Date 02.10.2003

> Male : 46 days Female : 0 days

**Duration of test** to gestation day 13-15

No. of generation

studies

Doses : 60, 120 and 210 ppm

Control group : ves NOAEL parental = 60 ppmother: NOAEL = 210 ppm

**Reproductive Toxicity** 

Method other Year 1985 **GLP** yes

**Test substance** as prescribed by 1.1 - 1.4

Result

: Exposure concentrations: The average mean daily analytical exposure concentrations (60.2, 120.4 and 208.9 ppm) were within 1% of the target levels (60, 120 and 210 ppm, respectively). Mean temperatures and humidities ranged from 22-25.5 degrees C and 24-27%, respectively.

Signs of toxicity: One of animals exposed to 210 ppm died after 2 days of exposure. On the previous day, this animal exhibited labored breathing, hypoactivity, poor control of the hind limbs, difficulty in standing, body tremors and involuntary movements. No unusual findings were observed at necropsy.

Body weights of males exposed to 210 ppm were approximately 6-9% lower than those of the control group during most of the exposure period. and remained lower than control (but were not significantly different) until the end of the study.

Animals exposed to 210 ppm exhibited signs of toxicity such as arched back (N = 8 on days 1-10, N = 3 on days 11-20 and 51-57, and N = 5 on days 41-50), hypoactivity (N = 12-15 at each 10-day interval up to day 50. and N = 4 from days 51-57), labored breathing (N = 10 on days 1-10, N = 3on days 11-20 and 31-40, N = 5 on days 21-30 and N = 1 on days 51-57), and salivation (N = 3 on days 1-10, and N = 10 - 12 at all other intervals). A few high dose animals (individual numbers were not stated) also exhibited abnormal behavior such as grinding of teeth, head bobbing, body tremors. involuntary movements, and pawing at the cage. A few of the animals exposed to 120 ppm exhibited salivation (N = 3-8 at all intervals) and hypoactivity (N = 3 at days 11-20). Signs of toxicity generally abated by the morning after exposure. Alopecia was observed in animals (N = 1 control, N = 2 low dose, N = 1 mid dose, N = 5 high dose) at one or more of their weekly physical examinations. No unusual treatment-related signs were observed in rats exposed to 60 ppm. The only remarkable finding at gross necropsy was a small right testis in one animal exposed to 120 ppm.

Fertility: There was no effect of treatment on male fertility. Efficiency of mating (34.4%, 30.6%, 29.8% and 27.1% in the control, low, mid and high dose groups) and pregnancy rate (90.5%, 97.6%, 90.0% and 97.4% in the respective groups) were comparable between groups. There was no difference in the numbers of live implants (ranged from 12.7 - 13.9), resorptions (ranged from 0.7 - 1.1), nidations (ranged from 13.8 - 14.9), corpora lutea (ranged from 13.1 - 15.2), preimplantation loss (4-8%) and

postimplantation loss (5-10%).

**Test condition** 

Animals: Virgin female Sprague Dawley rats (28 days old upon receipt) and virgin male Sprague Dawley rats (50 days old upon receipt) were acclimated for one week and examined for general health and the presence of pinworms and ectoparasites. Body weights of fifteen females and ten males that were taken upon receipt were 155-181 g and 80-103 g. respectively. No significant health problems were noted during the

acclimation period, and the animals were released for study. Males and females were individually caged (except during mating). Food and water were available ad libitum (except food was not available to males during exposure). Animals were housed in a room maintained at 72 +/- 2 degrees F and 40-60% relative humidity, under a 12 hr light/dark cycle.

Exposures: Exposures (6 hr/day, 5 days/week) occurred in 10 m3 Rochester-style stainless steel and glass inhalation chambers. A scheduled exposure day was cancelled due in clement weather. A new exposure day (exposure day 41) was used in its place. Due to inclement weather and building equipment failures, 2 exposures (days 33 and 43) were only for 4 hours and one exposure (day 32) was for 5 hours. Atmospheres of propionitrile vapor were generated using bubbler systems. The concentrations of material in the chambers were controlled by either adjusting the nitrogen flow though the propionitrile in the bubblers or by changing the amount of test material in the bubblers. Concentrations of test material in the chambers were measured 4 times daily (except during day 43) using a Miran 1A General Purpose Gas Analyzer. The nominal concentration of material, temperature and humidity also were measured (at intervals that were not listed).

Study conduct: Fifteen males per group were assigned to be exposed to target concentrations of 0, 60, 120 or 210 ppm test material. Exposure began when males were 43 days old. Mating was initiated when males and females were 16 and 12 weeks old, respectively. At this time, males had been 69 days on the study (which was sufficient to cover the spermatogenesis cycle of the rat), and had 46 days of exposure. Males were randomly mated (1: 1) with three untreated females (consecutively) that had been assigned to the corresponding treatment group (45 females were assigned per group). Exposure of males continued until the day after the last mating opportunity (57 exposure days). At night, after exposure, males were caged with their assigned female until copulation was confirmed (by presence of a copulatory plug or sperm in vaginal smear) or 5 nights without confirmed copulation. The day on which copulation was confirmed was considered gestation day 0.

After assignment to the study, males and females were weighed once per week. Mated females were weighed on gestation days 0 and 13. Males were given a thorough physical examination once per week and were observed for clinical signs of toxicity before and after exposures. All animals were checked twice daily for mortality and gross abnormalities (except for one day prior to mating when inclement weather permitted observations).

One half of the males of each group were euthanized on each of the 2 consecutive days at the end of the study. They had not been exposed to propionitrile for about 2 weeks prior to termination. Each male was given an external examination and weighed. The tissues and organs of the thoracic, scrotal and abdominal cavities were examined for gross lesions and the testes, epididymides, prostate glands and seminal vesicles were preserved in 10% neutral buffered formalin. Females that were not mated with males were euthanized and were not examined.

Mated females were euthanized on gestation day 13 (or the nearest workday up to gestation day 15). Females that were co-housed with males without confirmed copulation were euthanized during the second week after the last day of co-housing. Gross necropsies were performed on females that had copulated and those that had not. The tissues and organs of the thoracic and abdominal cavities were examined. Pregnancy status was determined, nidation sites and were classified, and corpora lutea were counted.

Statistical analyses: Body weight data were analyzed using Dunnett's test. Mating and pregnancy rate data were analyzed by a Fisher's exact test and an uncorrected chi-square test. Other data were analyzed by the Mann-Whitney U test. The Bonferroni inequality was assumed when comparing multiple treatments to control values for all tests except Dunnett's test. The

critical level for significance was p < 0.05.

**Test substance**: Purity of the test material was 96.1%. Impurities included acrylonitrile

(0.1%), adiponitrile (0.7%), p-nitrosodiphenylamine (0.08%) and water (< 0.1%). Analyses indicated no significant decomposition of the test material

over the course of the study.

**Conclusion**: There was no effect of treatment on fertility of males.

**Reliability** : (1) valid without restriction

Study is comparable to a guideline study.

Flag : Critical study for SIDS endpoint

07.08.2003 (25)

#### 5.8.2 DEVELOPMENTAL TOXICITY/TERATOGENICITY

Species : rat Sex : female

Strain : Sprague-Dawley

Route of admin. : gavage

**Exposure period** : Days 6 through 19 of gestation

Frequency of treatm. : daily

Duration of test: to Day 20 of gestationDoses: 20, 40, 80 mg/kg/dayControl group: yes, concurrent vehicle

NOAEL maternal tox. : = 40 mg/kg bw NOAEL teratogen. : = 80 mg/kg bw NOAEL Fetotoxicity : = 40 mg/kg bw NOAEL Embryotoxicity : = 40 mg/kg bw

Method: otherYear: 1981GLP: ves

**Test substance**: as prescribed by 1.1 - 1.4

**Remark**: Doses used in this study were chosen based on the results of a pilot study

(International Research and Developmental Corporation study IR-79-163, dated December 22, 1980) conducted with 5, 10, 20, 37.5 and 75

mg/kg/day. In this study, 75 mg/kg caused a moderate to severe decrease in maternal weight gain. Two out of 5 dams treated with 75 mg/kg/day also had an increased number of resorptions (16 early in one animal and 7 late

in the other).

In the main study, the number of fetuses and litters with sternebrae 5 and/or 6 unossified in the mid dose group (55 and 35, respectively) was "nearly comparable" to historical controls (3.8 - 23.7 in 13.0 - 73.9 litters). Since this was the only change observed in the mid dose fetuses, it is not considered to be indicative of fetotoxicity. Therefore, this dose was chosen as the NOAEL for fetotoxicity.

as the NOAEL for fetotoxicity.

The NOAEL for teratology was established by study personnel. The summary preparer established the NOAELs for maternal toxicity,

fetotoxicity and embryotoxicity.

Result : Maternal: One rat in the high dose group died of an undetermined cause

on gestation day 9. Survival was 100% in the other groups. There was a slight to moderate reduction in mean maternal body weight gain over the entire treatment period for the high dose group (118 g in treated vs. 134 g in control). This decrease was predominantly due to decreased body weight gain from days 6 to 9 (2 g in treated vs. 9 g in control). The mean

maternal adjusted body weight gain (body weight minus the uterus and contents) in the high dose group also was slightly reduced during the gestation period (52 g in treated vs. 60 g in control). Weights of animals in other treated groups were similar to controls.

There was no effect of treatment on the number of pregnancies, mean number of viable fetuses (all were viable), late resorptions, total implantations, or corpora lutea. There was a significant increase in the number of early resorptions (2.0 +/- 2.63 in treated vs. 0.7 +/- 0.81 in control) and a corresponding increase in the number of postimplantation losses in the high dose group. None of the animals aborted.

Fetal: There was no effect of treatment on fetal sex ratio. Average mean fetal body weight of fetuses from high dose animals were significantly less than controls (3.0 +/- 0.40 g in treated vs. 3.5 +/- 0.28 g in control). The number of control, 20 mg/kg/day, 40 mg/kg/day and 80 mg/kg/day fetuses (and litters) with malformations were 5 (3), 1 (1), 1 (1) and 0 (0), respectively. One fetus in the low dose group and one in the mid dose group had a diaphragmatic hernia. An increase in the number of fetuses and litters with sternebrae 5 and/or 6 unossified was noted in the mid dose group (55 and 35, respectively) and high dose group (92 and 66.7, respectively) when compared to the study control (21 and 14, respectively). An increase in the number of fetuses and litters with sternebrae 1 and/or 2, 3, and 4 unossified was also found in the high dose group (8 in 2 treated litters vs. 1 in control).

**Test condition** 

Rats: One hundred virgin female COBS CD rats (approximately 14 weeks old) were used in the study. They were acclimated for at least 10 days prior to mating. One female was mated with one male. Copulation was verified by the presence of a copulatory plug or sperm in a vaginal smear. The day that evidence of mating was detected was designated day 0 of gestation. Mated females were assigned in a block design to a vehicle control group and 3 treatment groups consisting of 25 rats each.

Test material: Dosing solutions of propionitrile were prepared daily at concentrations that permitted administration of 20, 40 and 80 mg/kg/day at a constant volume of 10 ml/kg. The material was dissolved in distilled water and shaken by hand to ensure dissolution. The test material was administered orally by gavage as a single daily dose on days 6 through 19 of gestation. The control group received distilled water at 10 ml/kg. Individual dosages were determined using body weights that were taken on gestation day 6.

Maternal observations: Prior to treatment, the dams were observed daily for mortality and overt changes in appearance and behavior. They were also observed daily from days 6 through 20 of gestation. Individual body weights were recorded on gestation days 0, 6, 9, 12, 16 and 20. On gestation day 20, all survivors were euthanized. The uterus was excised and weighed, and the number and location of viable and nonviable fetuses, early and late resorption and total number of implantations and corpora lutea were recorded. The abdominal and thoracic cavities and organs of the dams were examined grossly. Any dam that did not survive to scheduled termination also was necropsied. Maternal tissues were preserved when deemed necessary according to gross findings. Uteri from females that appeared nongravid were opened and placed in 10% ammonium sulfide solution for confirmation of pregnancy.

Fetal observations: All fetuses (total of 326, 302, 319 and 272 in the control, low, mid and high dose groups) were individually weighed and examined for external malformations and variations. Each fetus was sexed. Approximately one-half of the fetuses were placed in Bouin's fixative for subsequent visceral examination by razor-blade sectioning. The other fetuses were fixed in alcohol, macerated in potassium hydroxide and

stained with Alizarin Red S for subsequent skeletal examination.

Statistical analyses: The sex distribution and number of litters with malformations in treated animals were compared to controls using a Chisquare test with Yates' correction for 2 x 2 contingency tables and/or Fisher's exact probability test. The numbers of early and late resorptions and postimplantation losses were compared by the Mann-Whitney U-test. The mean number of viable fetuses, total implantations, corpora lutea and mean fetal body weights were compared by Bartlett's test for homogeneity of variance, analysis of variance (one-way), and the appropriate t-test (for equal or unequal variances). Dunnett's multiple comparison tables were used to judge significance of differences. The level of significance was p < 0.05.

**Test substance**: The purity of the test material was > 90%.

Reliability : (1) valid without restriction

Significant differences between weights and weight gains of treated dams

and controls were not designated.

Flag : Critical study for SIDS endpoint

10.08.2003 (22)

#### 5.8.3 TOXICITY TO REPRODUCTION, OTHER STUDIES

Type : other: examination of reproductive organs from 14 week study

Species : rat

Sex : male/female
Strain : Sprague-Dawley
Route of admin. : inhalation

**Exposure period** : 14 weeks (total of 63 exposure days)

**Frequency of treatm.** : 6 hours/day, 5 days per week

Post exposure period

**Doses** : 60, 120, 209 ppm

Control group : yes

 NOAEL
 : < 60 ppm</td>

 LOAEL
 : = 60 ppm

 Method
 : other

 Year
 : 1984

 GLP
 : yes

**Test substance** : as prescribed by 1.1 - 1.4

Remark : The NOAEL listed above is for reproductive effects. The NOAEL for other

systemic effects was < 60 ppm. Additional information about systemic

toxicity is listed in Section 5.4.

Result : Test material concentrations: The nominal concentrations (+/- SD) were

56.0 +/- 8.8, 119.1 +/- 16.9 and 203.0 +/- 19.6. Corresponding analytical concentrations were 60.2 +/- 1.0, 120.3 +/- 1.1 and 209.0 +/- 1.3 ppm.

Terminal absolute and relative testes weights after 14-weeks treatment are listed in the following table:

	Testis, mean absolute weight (g)		Testis, mean weight relative to body weight		
Group	Left	Right	Left	Right	
Control	2.618	2.638	0.572	0.575	
Low	2.437	2.482	0.511*	0.521*	
Mid	2.470	2.539	0.575	0.592	
High	2.302*	2.330	0.589	0.595	

<sup>\*</sup> Significantly different from control (p < 0.05).

Absolute left testes weights were decreased in males exposed to 209 ppm

and weight-adjusted (relative) testicular weights were decreased at 60 ppm. Since the effects were not consistent and/or dose-related, the effects on testicular weights do not appear to be related to treatment. No treatment-related histopathological changes were noted in the ovaries, testes (with epidiymides), pituitary, prostate, uterus (with cervix), mammary gland or thyroid.

#### **Test condition**

Animals: Rats were acclimated for at least 10 days prior to use. Two days prior to the start of the study, males weighed 174-200 g and females weighed 132-145 g. On the first day of the study, the animals were 43 days old. Animals were randomly allocated by body weight into 4 groups of 15 animals/sex/group. Animals were individually housed in suspended mesh cages and given food and water ad libitum (except during exposure). Animal rooms were maintained at 70-74 degrees C and 35-60% relative humidity, with a 12 hour light/dark cycle.

Exposure conditions: Exposures (6 hr/day, 5 days/week) occurred in 10 m3 Rochester-style stainless steel and glass inhalation chambers. Rats were placed individually in wire mesh cages that were suspended in the chambers by 3-tiered racks. Males were placed on one side and females on the other. The concentrations of material in the chambers (20, 120 or 210 ppm) were controlled by either adjusting the nitrogen flow though the propionitrile in the bubblers or by changing the amount of test material in the bubblers. The bubbler was connected to a side port in the vertical particle-size separator, which was in turn connected to the air inlet at the top of the inhalation chamber. One bubbler was used in each of the generation systems. Airflow was maintained at a constant flow of 1727 liters/min. Nominal concentration measurements were determined daily for each chamber following exposure, by dividing the amount of test material delivered to the chamber (the difference between the pre- and postexposure weights) over the 6-hr exposure period by the total air volume during the same period. Concentrations of test material in the chambers were measured 4 times daily using a Miran 1A General Purpose Gas Analyzer. Additional samples of atmosphere from 9 specified locations in each chamber were also taken at 3 different times to determine if the vapor was distributed uniformly.

Test conduct: Animals were observed for clinical signs between the second and fifth hour of each exposure. Estimations of the percentages of animals exhibiting hypoactivity, eye irritation and breathing difficulties were made. All animals were individually examined for gross signs of toxicity preceding and following each exposure and checked for mortality. Each animal was weighed and given a thorough examination for gross signs of toxicity on a weekly basis.

Animals were euthanized after 14 total weeks on the study. Terminal body weights were obtained (following an overnight fast). Detailed necropsies were conducted on all rats that died during the course of the study, those that were killed moribund, and those that survived to study termination. The testes (with epididymides) were weighed. The following tissues were fixed in 10% neutral formalin: ovaries, mammary gland, thyroid, pituitary, prostate, testes (with epididymides) and uterus (with cervix). Tissues were processed, embedded in paraffin, cut at five microns, stained with hematoxylin and examined microscopically.

Statistical analyses: Organ weight data were analyzed using Dunnett's test. Organ to body weight ratios were analyzed using the Mann-Whitney test, with the Bonferroni inequality. Data for frequencies of microscopic lesions were evaluated with the Fisher's exact test with the Bonferroni inequality.

Test substance Reliability

- : The purity of the test material was 96%. Impurities were not listed.
- : (2) valid with restrictions

The study is comparable to a guideline study.

10.08.2003 (34)

#### 5.9 SPECIFIC INVESTIGATIONS

**Endpoint**: Mechanistic Studies

Study descr. in chapter : Reference : Type :

Species: mouseSex: maleStrain: CD-1Route of admin.: i.p.

No. of animals :

Method: otherYear: 1981GLP: no data

**Test substance** : as prescribed by 1.1 - 1.4

**Remark**: n-Butyronitrile also was tested in this study. The results with this material

were similar to those of propionitrile.

This study is considered to be valid without restriction. The study was

conducted and documented in a thorough manner.

**Result** : In the first study, the mortality rate for animals treated only with 45 mg/kg

propionitrile was 9/10. Co-treatment with sodium nitrite reduced the rate to 5/9. None of the 10 animals that were co-treated with sodium thiosulfate

died.

In the carbon tetrachloride study, the mortality rate for animals treated only with 45 mg/kg propionitrile was 8/10. None of the animals co-treated with

carbon tetrachloride died.

Cyanide concentrations in liver and brain of mice co-treated with sodium thiosulfate or carbon tetrachloride were significantly less than those of mice treated with propionitrile alone. In mice receiving propionitrile only, 26.7 +/-8.0 (mean +/- SD) and 12.8 +/- 5.4 nmol/g cyanide were found in liver and brain, respectively. In mice receiving propionitrile plus sodium thiosulfate, 2.0 +/- 0.7 and 0.8 +/- 0.8 nmol/g cyanide were found in liver and brain, respectively. In mice receiving propionitrile plus carbon tetrachloride, 0.9 +/- 1.1 and 1.5 +/- 1.4 nmol/g cyanide were found in liver and brain,

respectively.

**Test condition**: Male CD mice (30 g) were divided into 3 groups of ten animals each. One

group received 45 mg/kg i.p. propionitrile only, another received i.p. injections of 75 mg/kg sodium nitrite (a cyanide antagonist) 20 minutes before and 100 minutes after i.p. injection of 45 mg/kg propionitrile, and another received i.p. injections of 1 g/kg sodium thiosulfate (a cyanide antagonist) 20 minutes before and 80 and 180 minutes after i.p. injection of

45 mg/kg propionitrile.

Two other groups of 10 mice received either 0.2 ml of vegetable oil or 0.2 ml of 20% carbon tetrachloride (a hepatotoxic dose) in vegetable oil subcutaneously, 24 hours before i.p. treatment with 45 mg/kg propionitrile.

In both experiments, animals were observed for 7 days. Mortality data were analyzed statistically by the chi-square test. The criterion for significance was p < 0.05.

In an additional study, the concentrations of cyanide in liver and brain were determined in a) 5 mice treated only with 28 mg/kg propionitrile (i.p.), b) 5

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> mice given 1 g/kg sodium thiosulfate 20 minutes before and 80 minutes after 28 mg/kg propionitrile (i.p.), and c) 5 mice given 0.2 ml of 20% carbon tetrachloride subcutaneously 24 hours before i.p. treatment with 28 mg/kg propionitrile. All mice were killed 2.5 hours after propionitrile injection (if still alive at this time). The livers and brains were excised as soon as possible after death, quick-frozen and weighed. Cyanide concentrations were determined by the method of Bruce et al. (Anal Chem 27: 1346-1347,

1955). Results were analyzed using an unpaired t-test. Test substance

: The purity of the test material was 99%. No free cyanide was found in

solutions made in distilled, deionized water.

Conclusion : Propionitrile is activated by the liver to release cyanide, which is

responsible for acute toxicity.

05.08.2003 (35)

#### 5.10 EXPOSURE EXPERIENCE

#### 5.11 ADDITIONAL REMARKS

6. Analyt. Meth. for Detection and Identification	107-12-0 02.10.2003
6.1 ANALYTICAL METHODS	
6.2 DETECTION AND IDENTIFICATION	
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7. Eff. Against Target Org. and Intended Uses			<b>Id</b> 107-12-0	
		Date	02.10.2003	
7.1	FUNCTION			
7.2	EFFECTS ON ORGANISMS TO BE CONTROLLED			
7.3	ORGANISMS TO BE PROTECTED			
7.4	USER			
7.5	RESISTANCE			

# **Id** 107-12-0 8. Meas. Nec. to Prot. Man, Animals, Environment **Date** 02.10.2003 8.1 METHODS HANDLING AND STORING 8.2 FIRE GUIDANCE 8.3 EMERGENCY MEASURES 8.4 POSSIB. OF RENDERING SUBST. HARMLESS 8.5 WASTE MANAGEMENT SIDE-EFFECTS DETECTION 8.6 8.7 SUBSTANCE REGISTERED AS DANGEROUS FOR GROUND WATER 8.8 REACTIVITY TOWARDS CONTAINER MATERIAL

9. References Id 107-12-0
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# 10. Summary and Evaluation **Id** 107-12-0 **Date** 02.10.2003 10.1 END POINT SUMMARY 10.2 HAZARD SUMMARY 10.3 RISK ASSESSMENT